



# Radical Protein Footprinting in Stabilized Whole Blood

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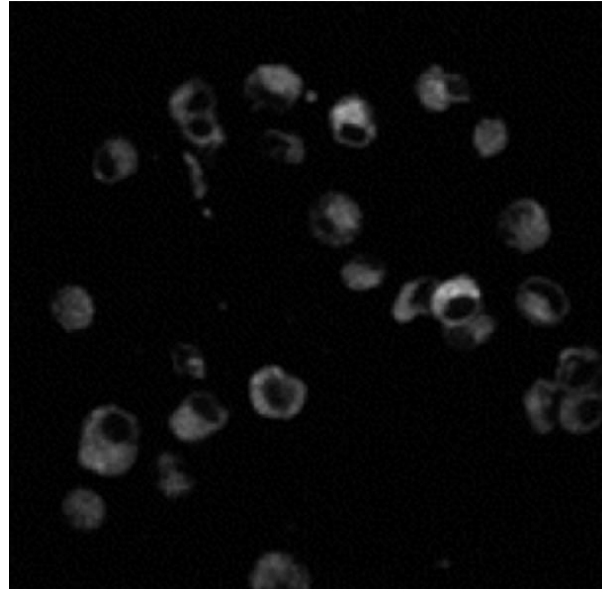
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FCOI Statement: J.S.S. and L.M.J. disclose a significant interest in GenNext Technologies, Inc., a growth-stage company seeking to commercialize benchtop HRPD to support the pharmaceutical industry

# Structural Proteomics by FPOP: From Test Tubes to Nematodes

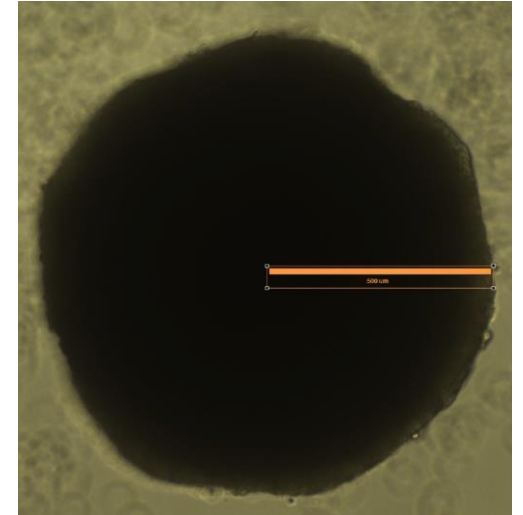
- Hydroxyl radical protein footprinting (HRPF) has long history *in vitro*
- In 2015, Espino Mali and Jones published the first example of FPOP in live cells
- Since been expanded to live nematodes and 3D cell cultures
- Mammalian tissues still out of reach due to strong tissue UV absorbance issues

Cultured Cells



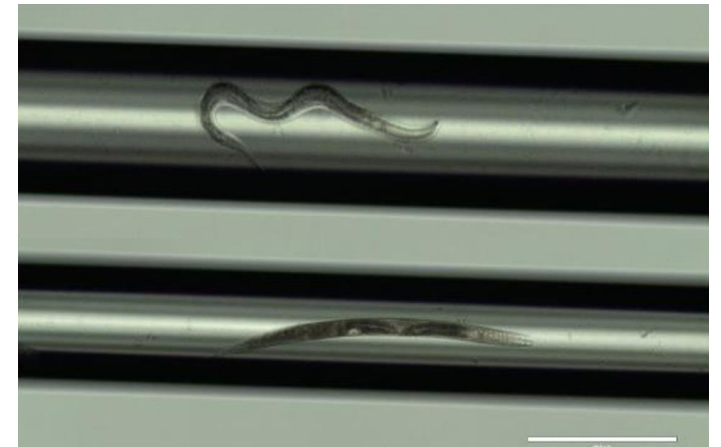
*Anal. Chem.* 2015, 87, 7971–7978

Solid Tumor Spheroids



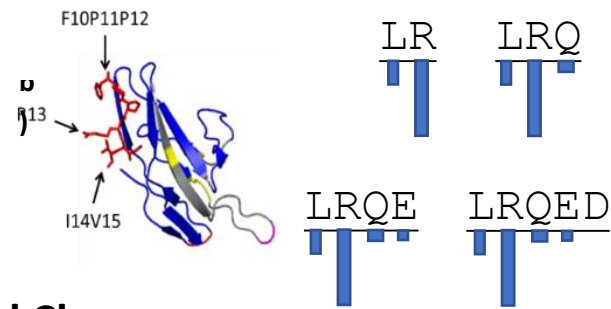
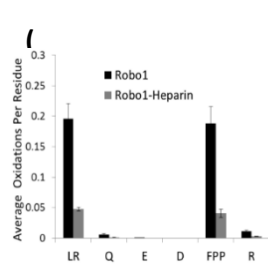
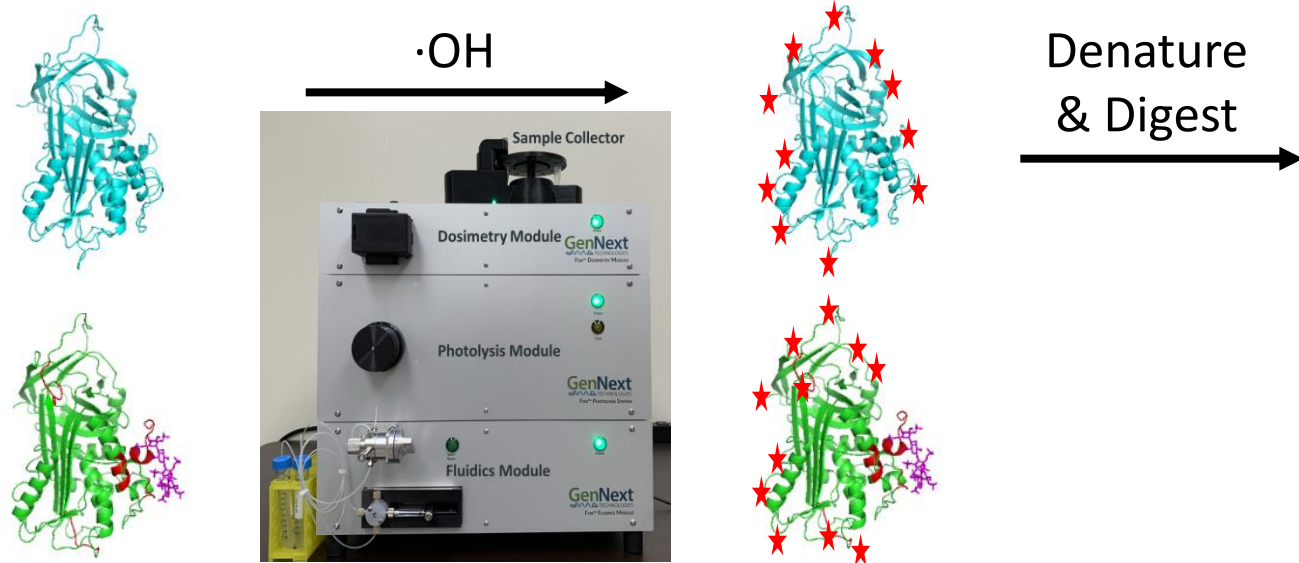
*J. Am. Soc. Mass Spectrom.* 2023, 34, 3, 417–425

Nematodes

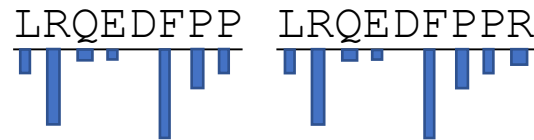


*Anal. Chem.* 2019, 91, 10, 6577–6584

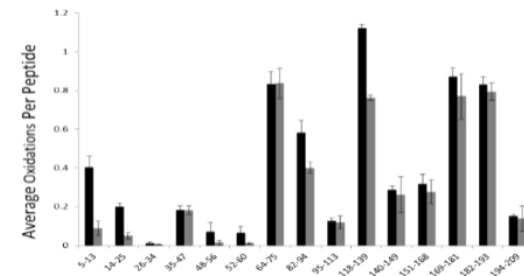
# General HRPF Workflow



**Amino Acid-Level Changes**  
**Structural Models**

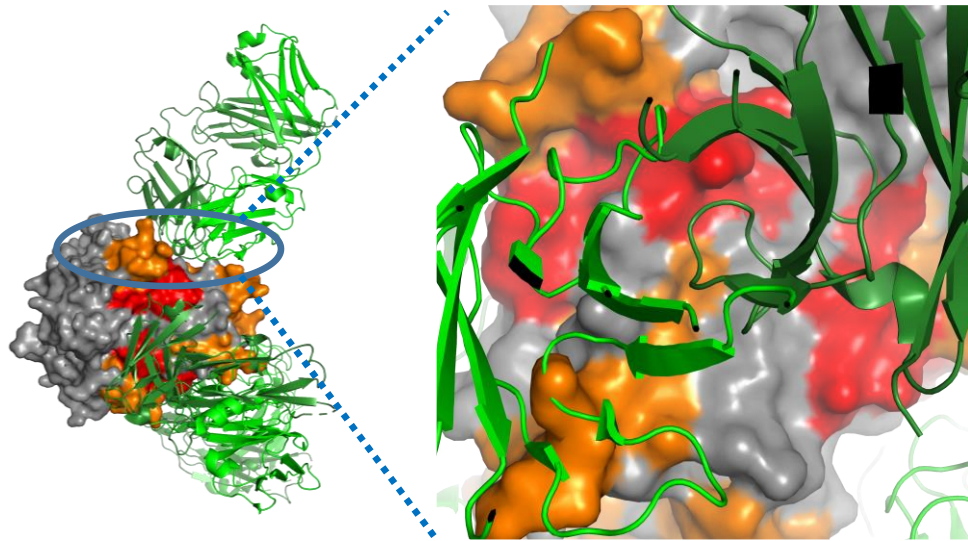


LC-MS



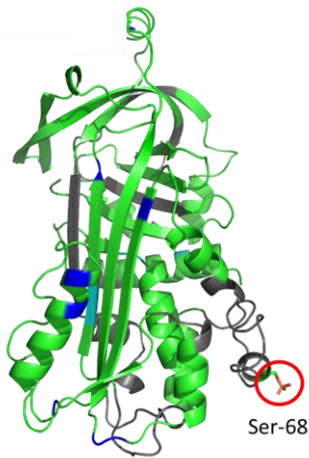
**Peptide-Level Changes**

# High Resolution Epitope Mapping

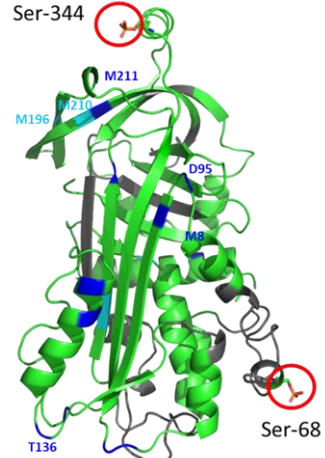


# Structural Impacts of PTMs

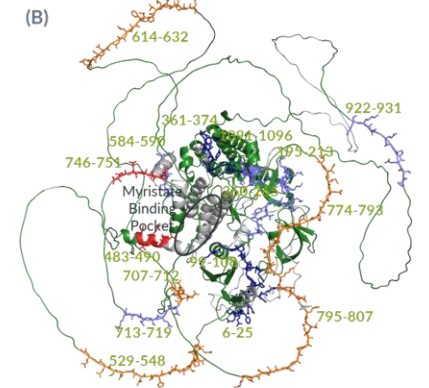
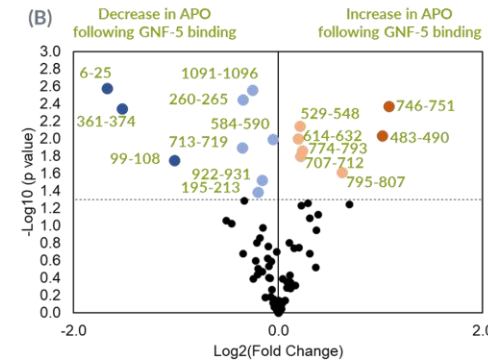
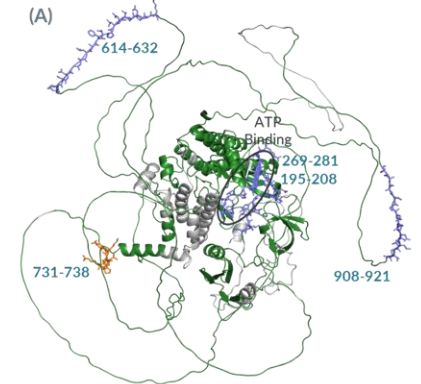
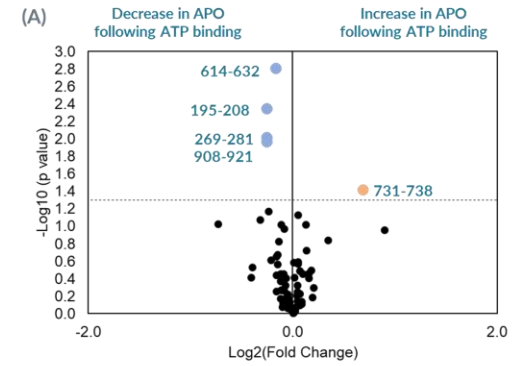
Non-PO3 Vs. Mono-PO3



Non-PO3 Vs. Di-PO3



# Allosteric Modulators



# In-Blood FPOP

- Major potential impact in structural pharmacology
  - Anti-drug antibody interactions
  - Drug:target interactions
  - Drug:off-target interactions
  - Post-administration aggregation
- Major potential for structural proteomics in human fluids
  - Liquid tumor analysis
  - Biology and diseases of blood, lymph, CSF, urine, etc.

## Anti-drug antibody prevalence in oncology checkpoint inhibitor therapy

| mAb                    | EMA first approval | Structure        | Isotype      | Target | ADAs %    |
|------------------------|--------------------|------------------|--------------|--------|-----------|
| Atezolizumab           | 2017               | Humanized        | IgG1, kappa  | PD-L1  | 30–54.1   |
| Avelumab               | 2017               | Fully human      | IgG1, lambda | PD-L1  | 4.1–5.9   |
| Cemiplimab             | 2019               | Fully human      | IgG4, kappa  | PD-1   | 1.3       |
| Durvalumab             | 2018               | Engineered human | IgG1, kappa  | PD-L1  | 1.7–6.6   |
| Nivolumab              | 2015               | Fully human      | IgG4, kappa  | PD-1   | 4.1–37.8  |
| Pembrolizumab          | 2015               | Humanized        | IgG4, kappa  | PD-1   | 0.7–2.5   |
| Ipilimumab             | 2011               | Fully human      | IgG1, kappa  | CTLA-4 | 1.1–26    |
| Nivolumab + ipilimumab | –                  | –                | –            | –      | 23.8–37.8 |

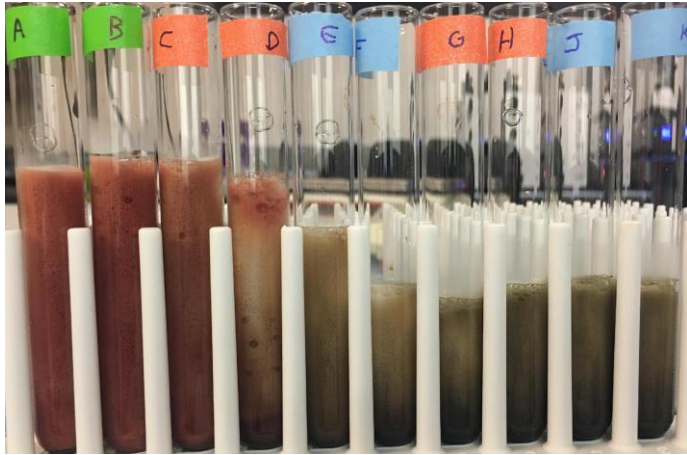
# Catalase Activity and Inhibition

500  $\mu\text{L}$  blood  
80  $\mu\text{L}$  30%  $\text{H}_2\text{O}_2$

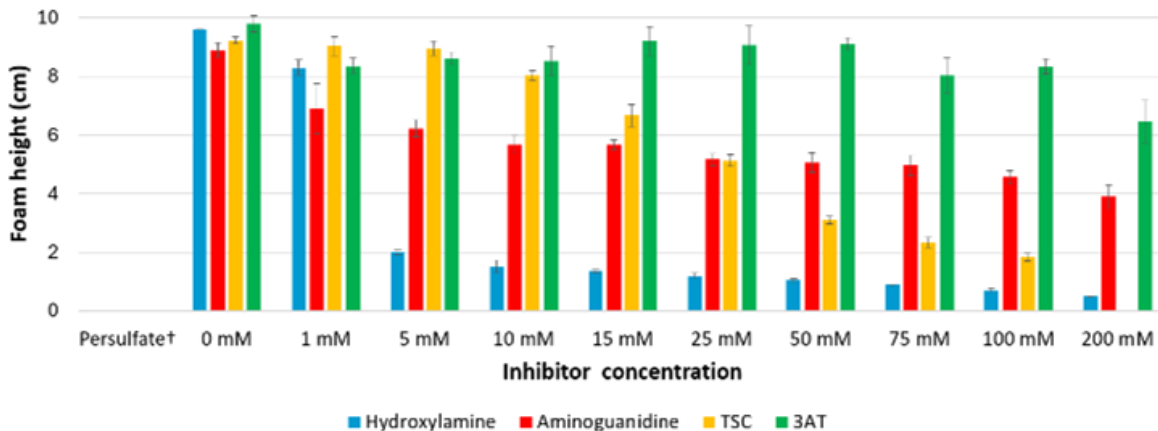
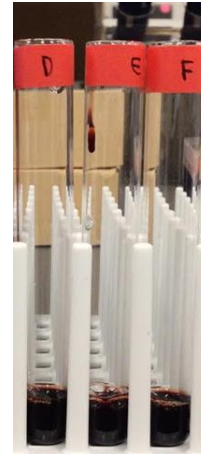
500  $\mu\text{L}$  blood  
80  $\mu\text{L}$  2.5M  $\text{Na}_2\text{S}_2\text{O}_8$

5 mM HA    100 mM HA

No inh.    25 mM HA    142 mM HA

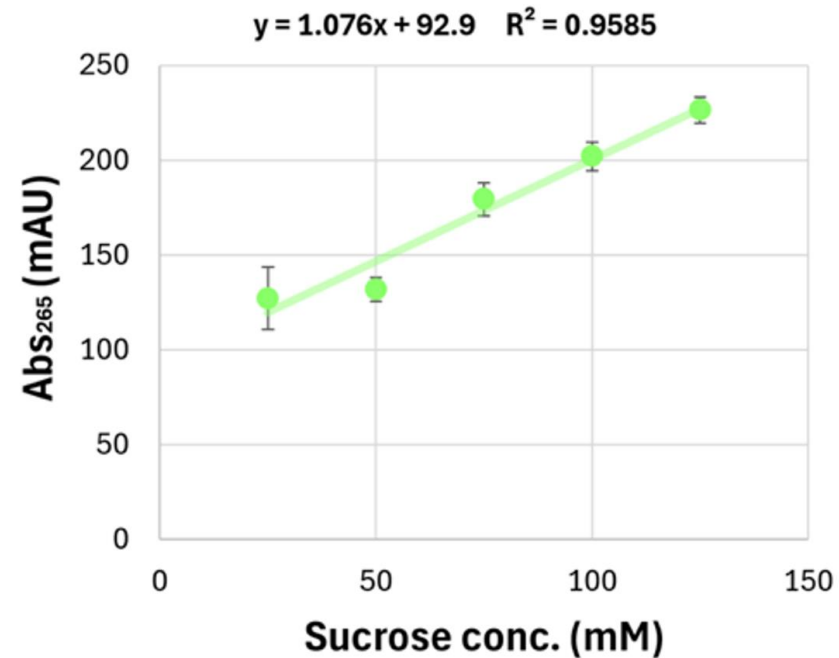
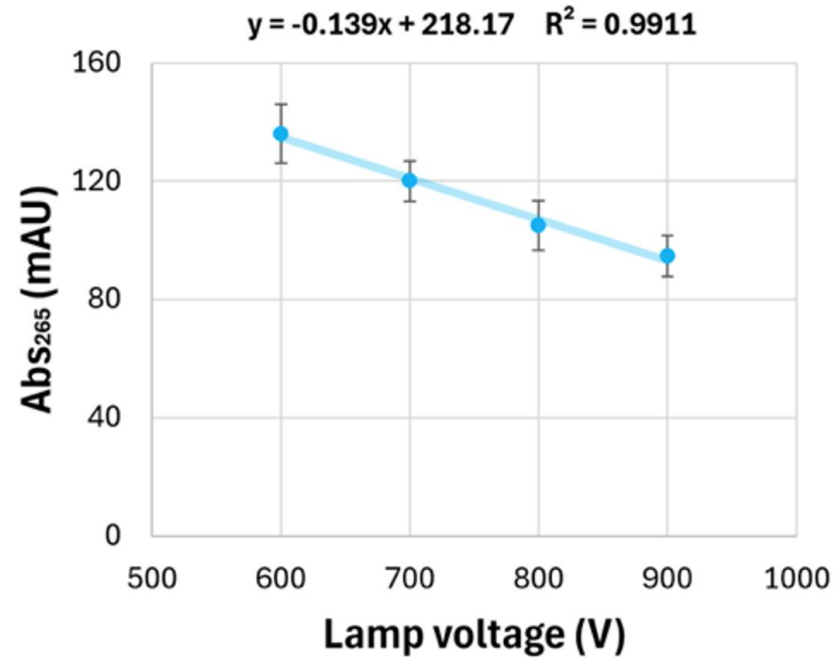


No inh.



- Small amount of mild detergent captures bubbles made by catalase
- No amount of quencher tested completely inhibited blood catalase
- High concentrations of inhibitor changed blood characteristics markedly
- No measurable gas production with persulfate

# In Vitro Persulfate Oxidation in Fox<sup>®</sup> System

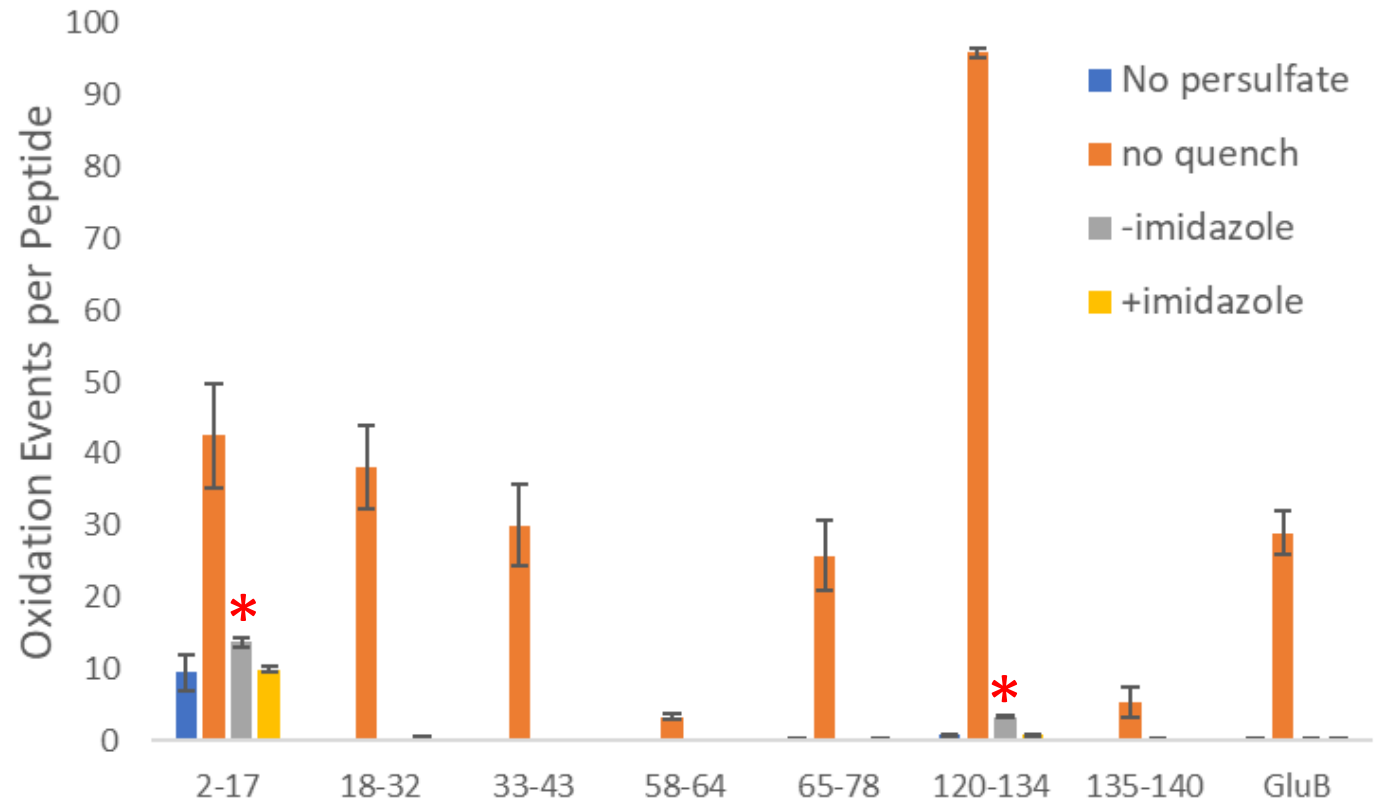


- Fox Photolysis System can efficiently photoactivate persulfate to oxidize proteins, peptides and small molecules
- Real-time adenine dosimetry works *in vitro*; absorbance decreases as more radical is created, decreases as more radical is scavenged

# New Persulfate Quench System

- Peroxide-based FPOP quenching with DMTU and methionine still had some peptides with elevated background oxidation
- Addition of 200 mM imidazole to quench eliminated background oxidation in these peptides
- Hydroquinone and GSH also seem to work, but have other issues

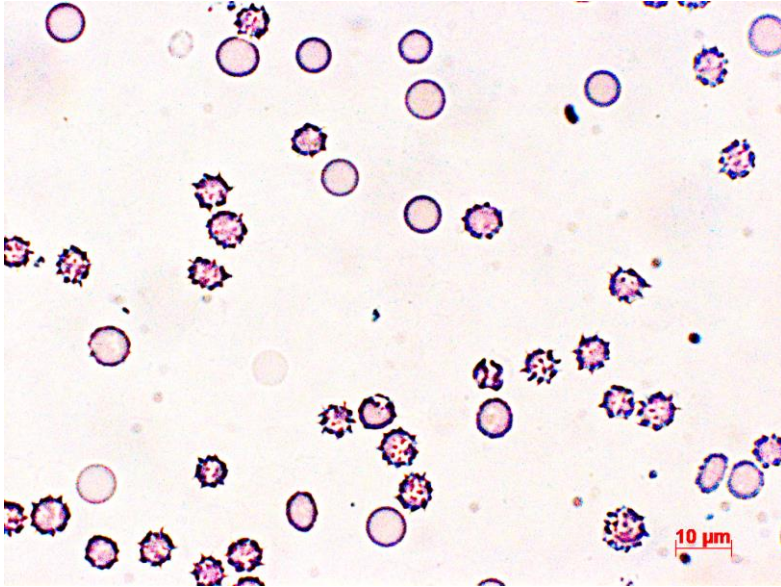
Protein in quench, 100 mM persulfate, 900V lamp  
Quench: 200 mM DMTU, 70 mM methionine,  $\pm$  200 mM imidazole



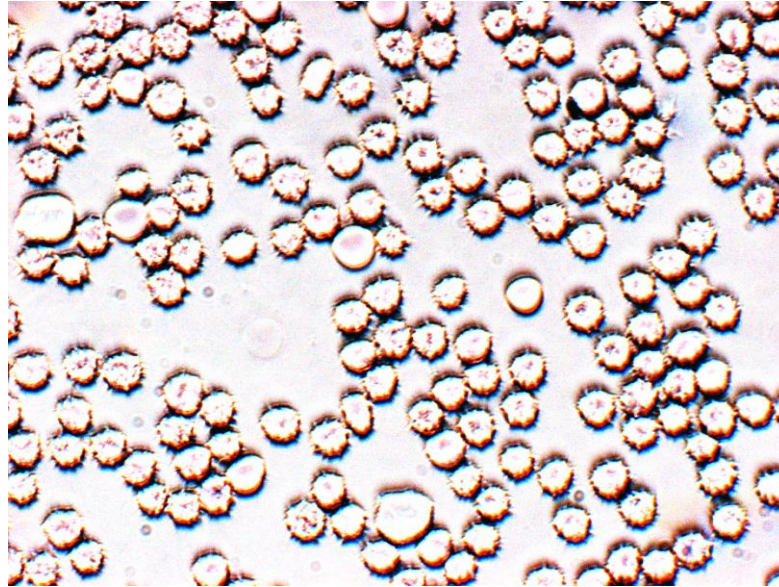
\* = significant increase from no persulfate to -imidazole quench



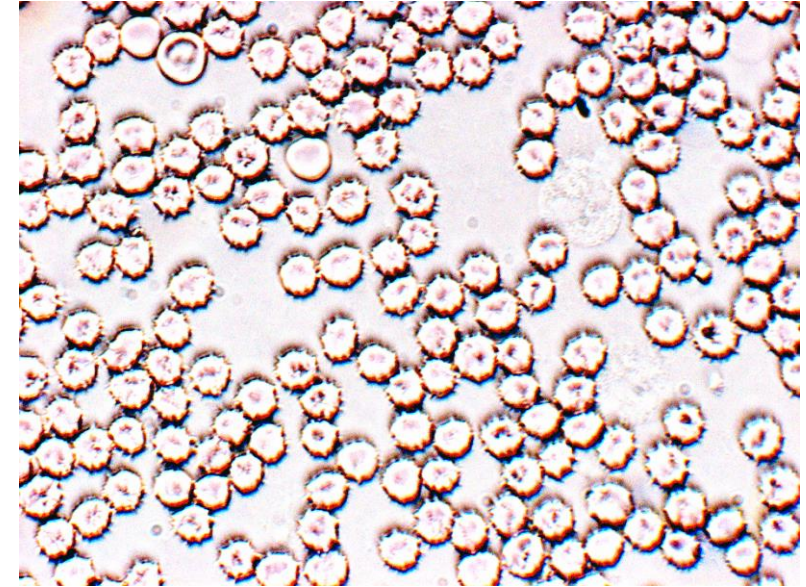
# Blood Cell Morphology in 200 mM Sodium Persulfate



Canine Blood



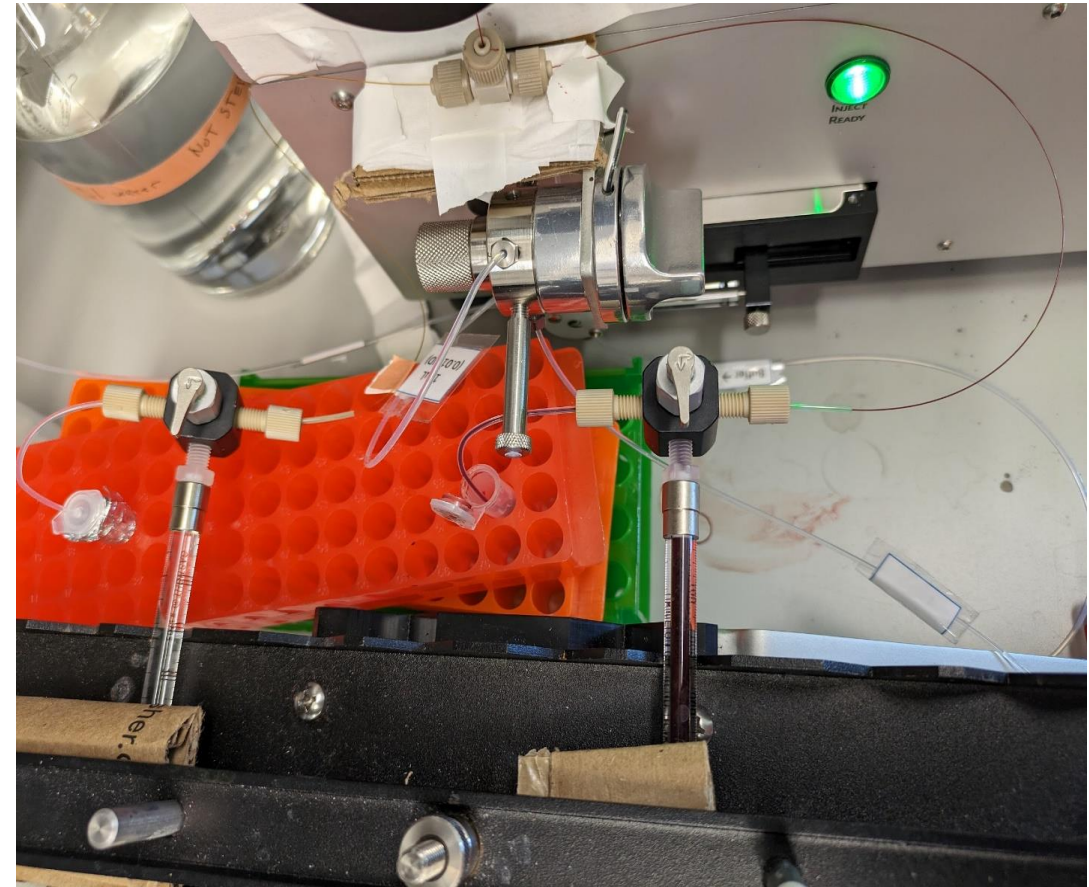
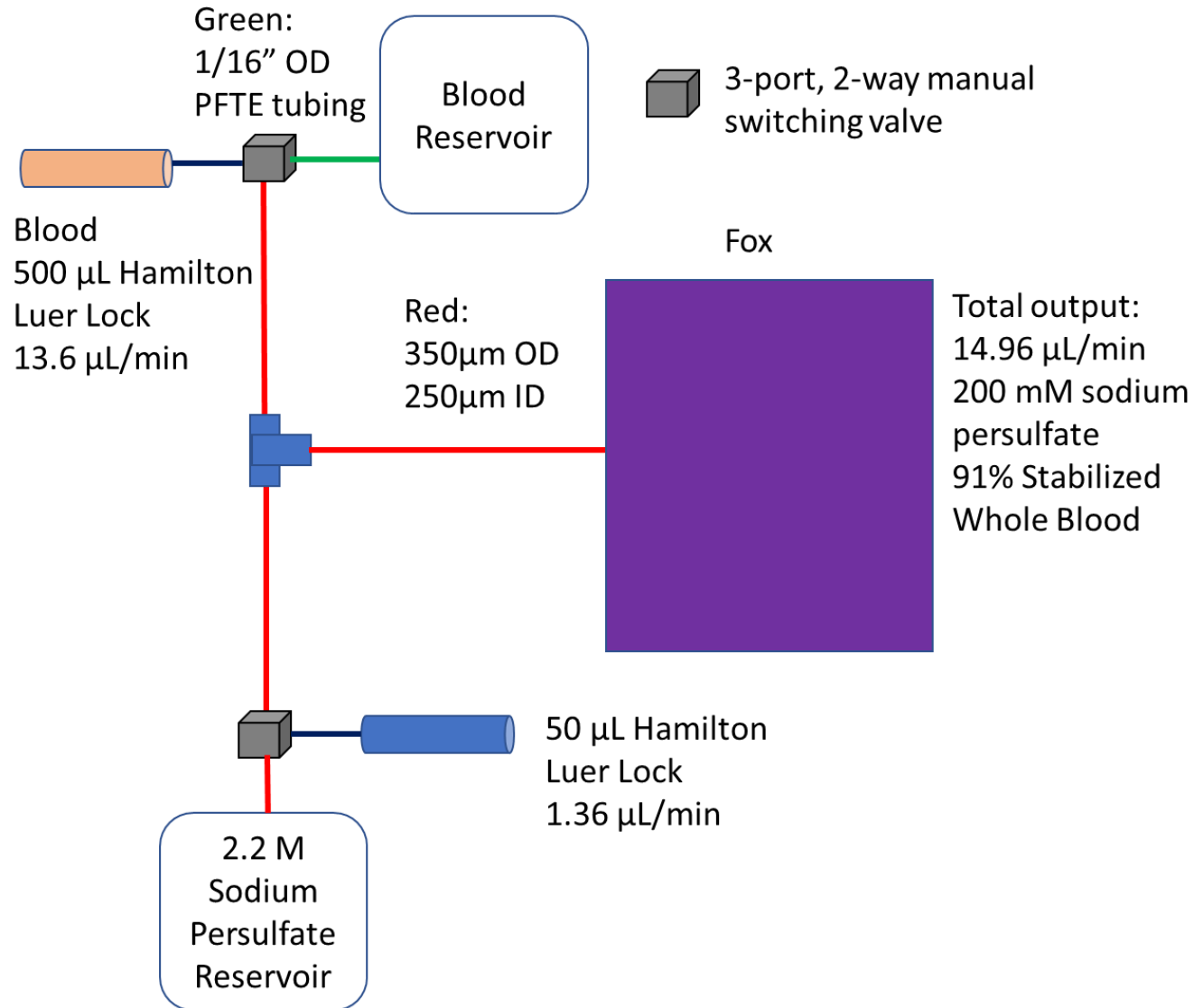
+200 mM sodium persulfate



+200 mM sodium chloride

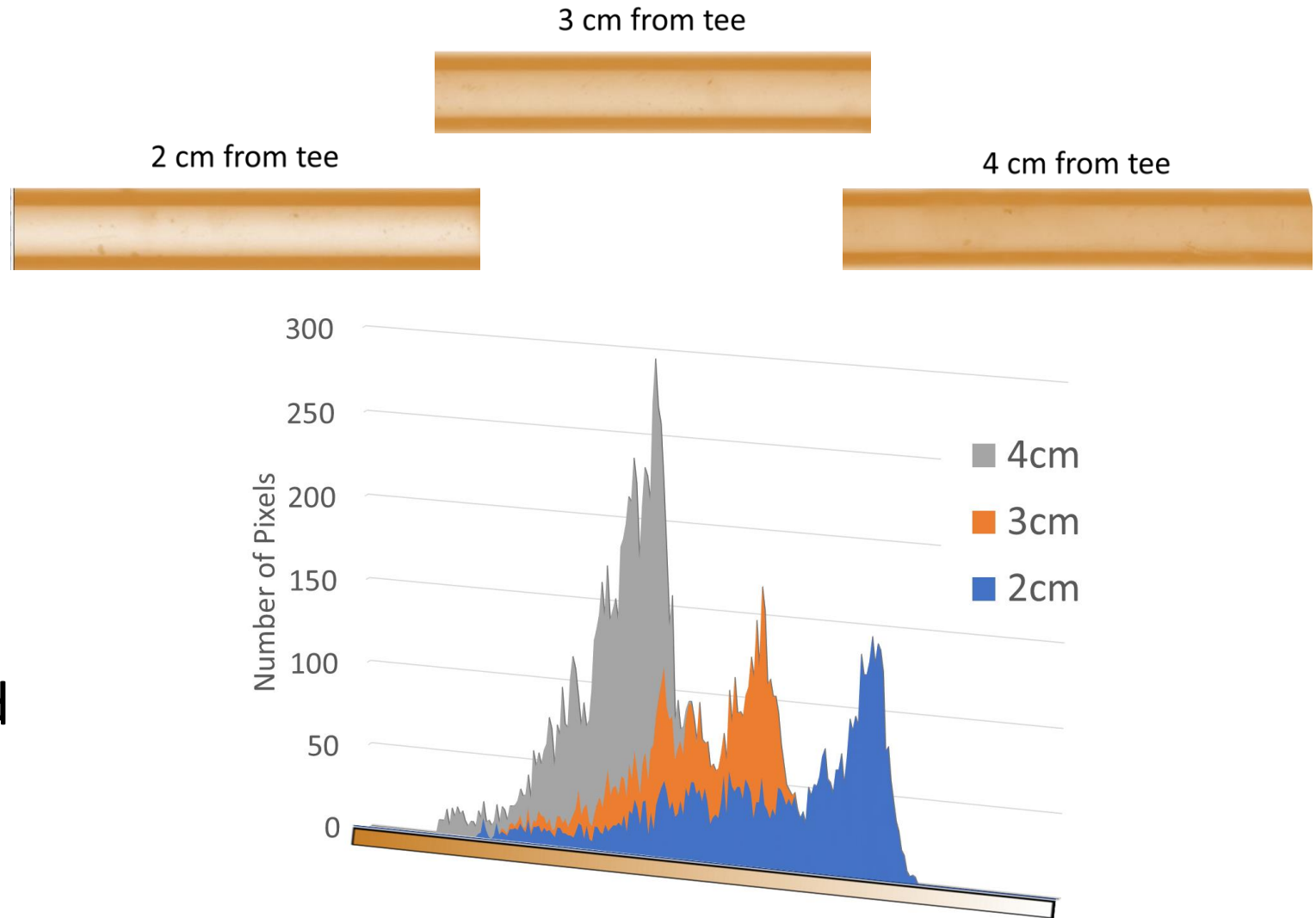
- No increase in ghost cells observed
- Moderate increase in hypertonicity observed in In-Blood RPF conditions
- Gross morphological changes indistinguishable from addition of equivalent concentration of sodium chloride on footprinting timescale (<1 minute)

# In-Blood FPOP System Design



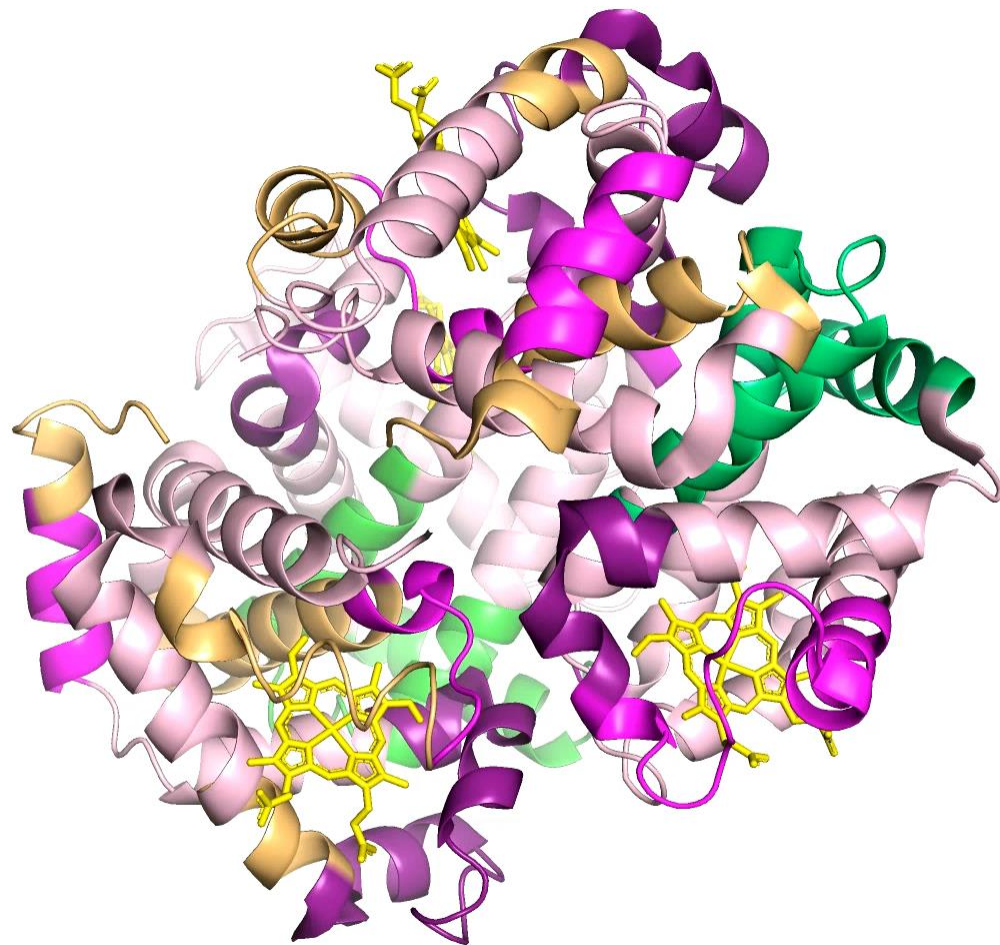
# Persulfate Mixing: Dye Imaging Test

- Two dyes: Red ( $1.3 \mu\text{L}/\text{min}$ ) and Blue ( $13 \mu\text{L}/\text{min}$ )
- Ran each dye by itself and measured in capillary to set color RGB values
- Set RGB values for two dyes in ImageJ Colour Deconvolution2 plugin
- Measured distribution of red dye across cross-section of capillary at different distances from mixing tee



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**Hemoglobin**  
**900V Flash Voltage**



**Green: Alpha-globin**  
**regions detected not**  
**oxidized**  
**Lt. Orange: Beta-globin**  
**regions detected not**  
**oxidized**  
**Yellow: Heme**

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Avg. Oxidations per Peptide

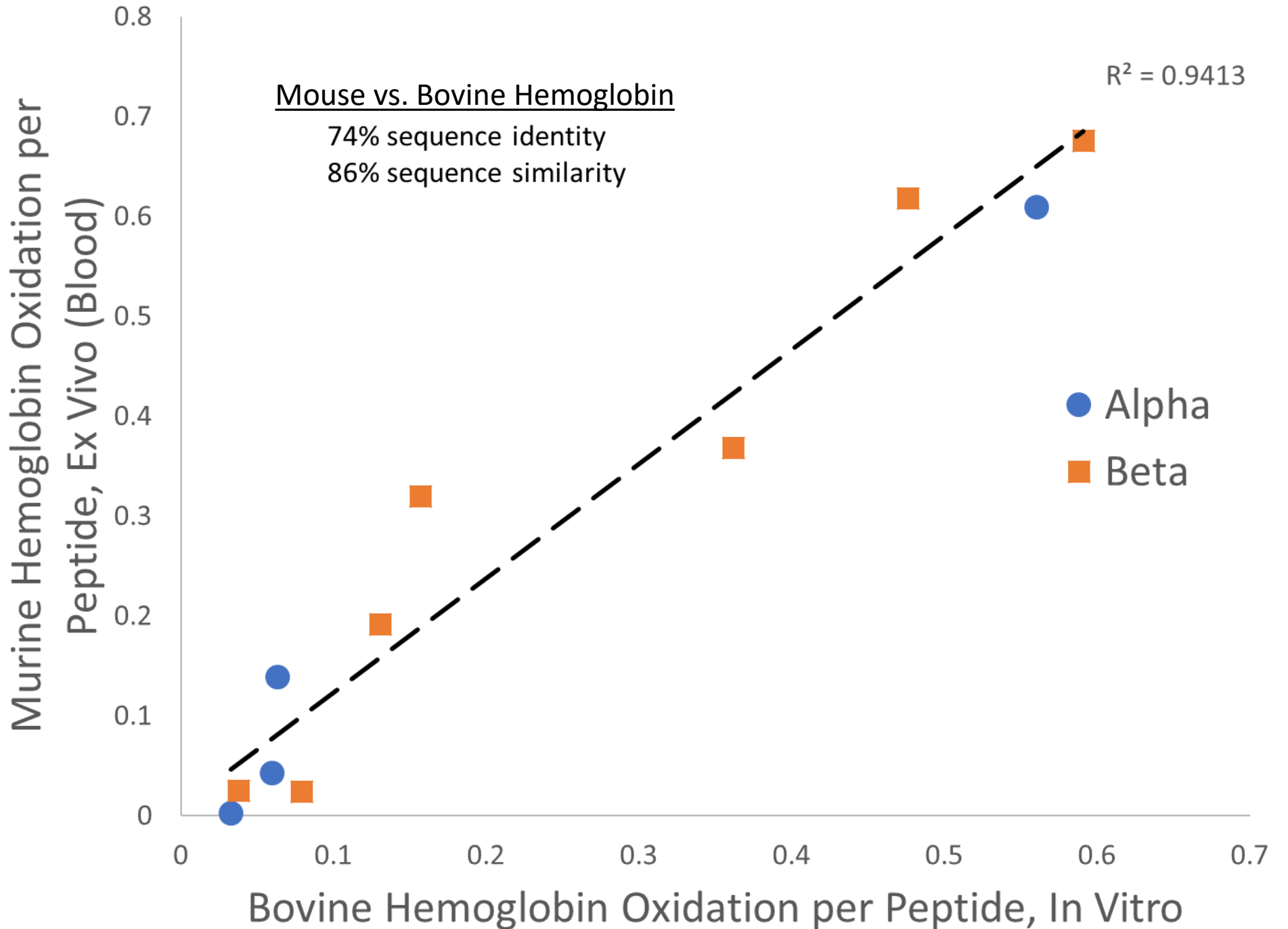


### In Blood

- 200 mM persulfate
- 900V lamp
- Murine blood

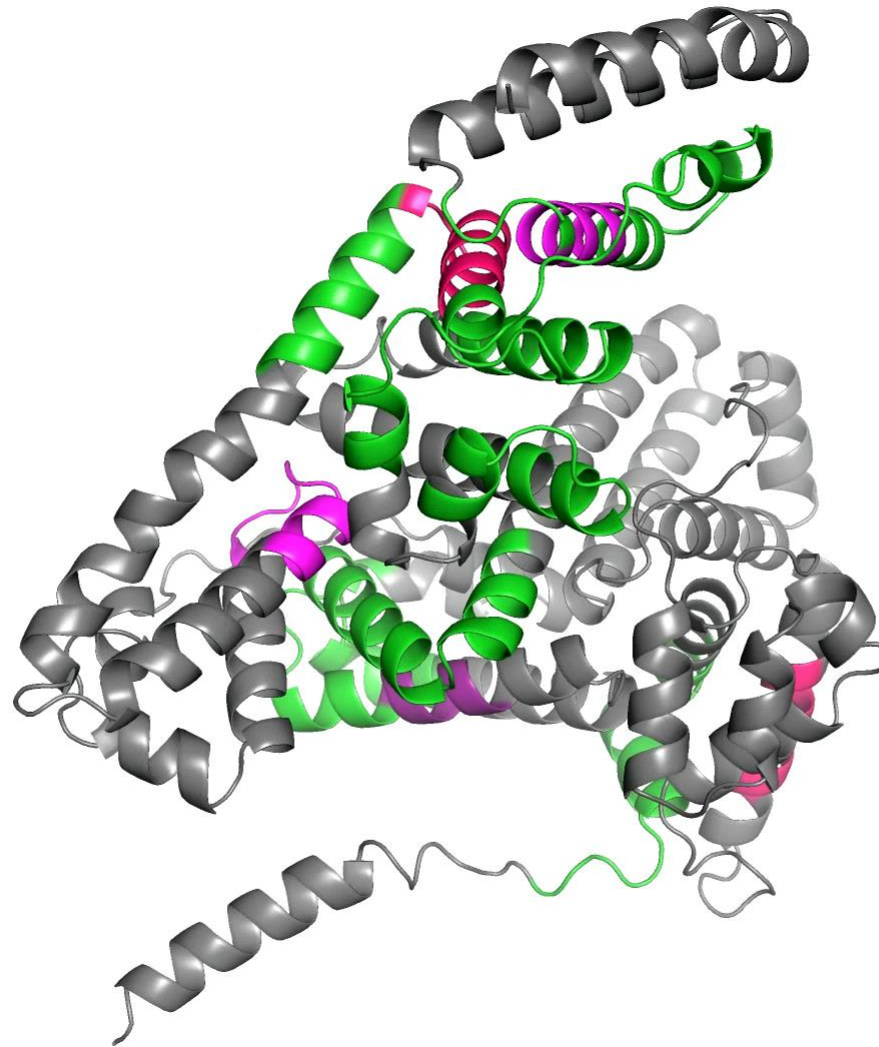
### In Vitro

- 100 mM persulfate
- 900V lamp
- Bovine hemoglobin



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Albumin  
900V Flash Voltage



**Green: Albumin regions  
detected not oxidized**  
Gray: Albumin regions  
not detected

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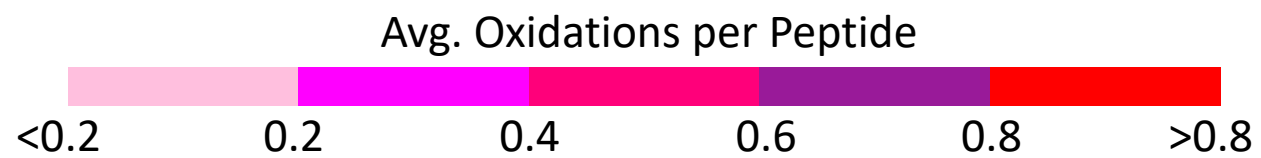
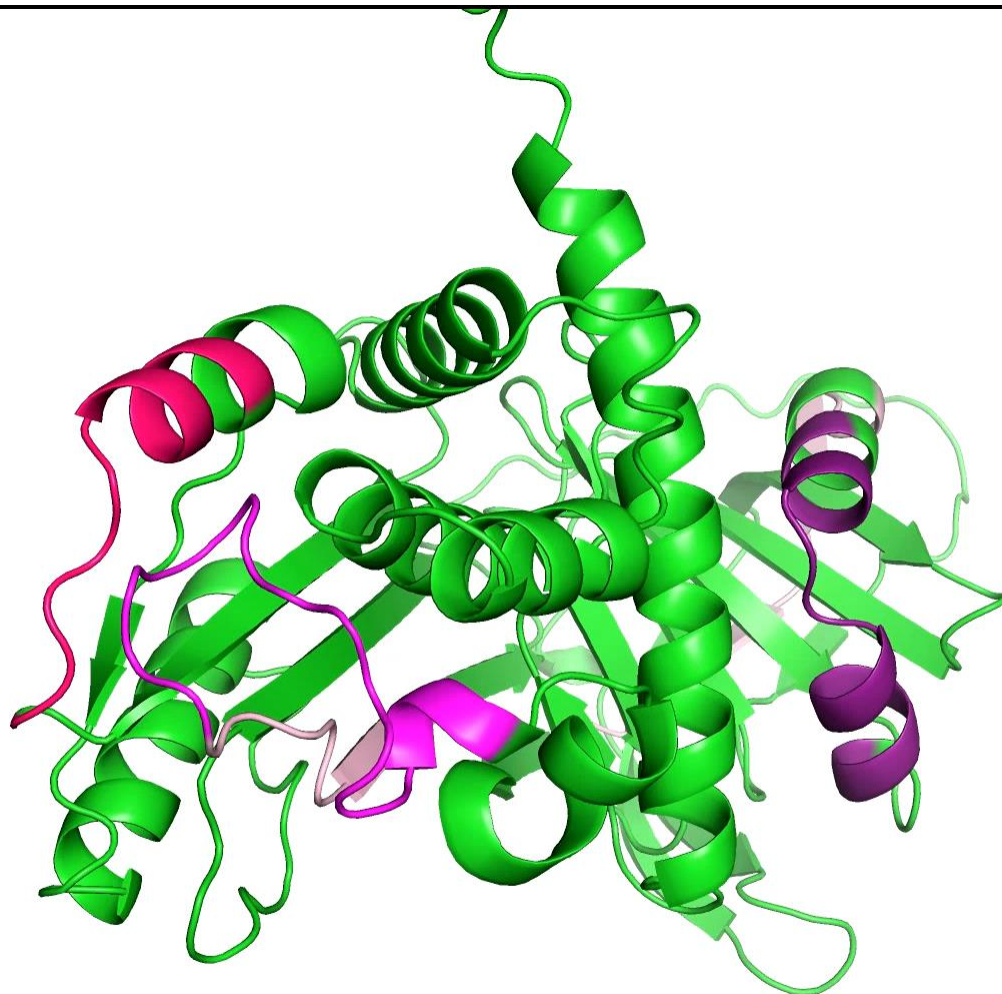
Avg. Oxidations per Peptide



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Serine protease inhibitor  
900V Flash Voltage

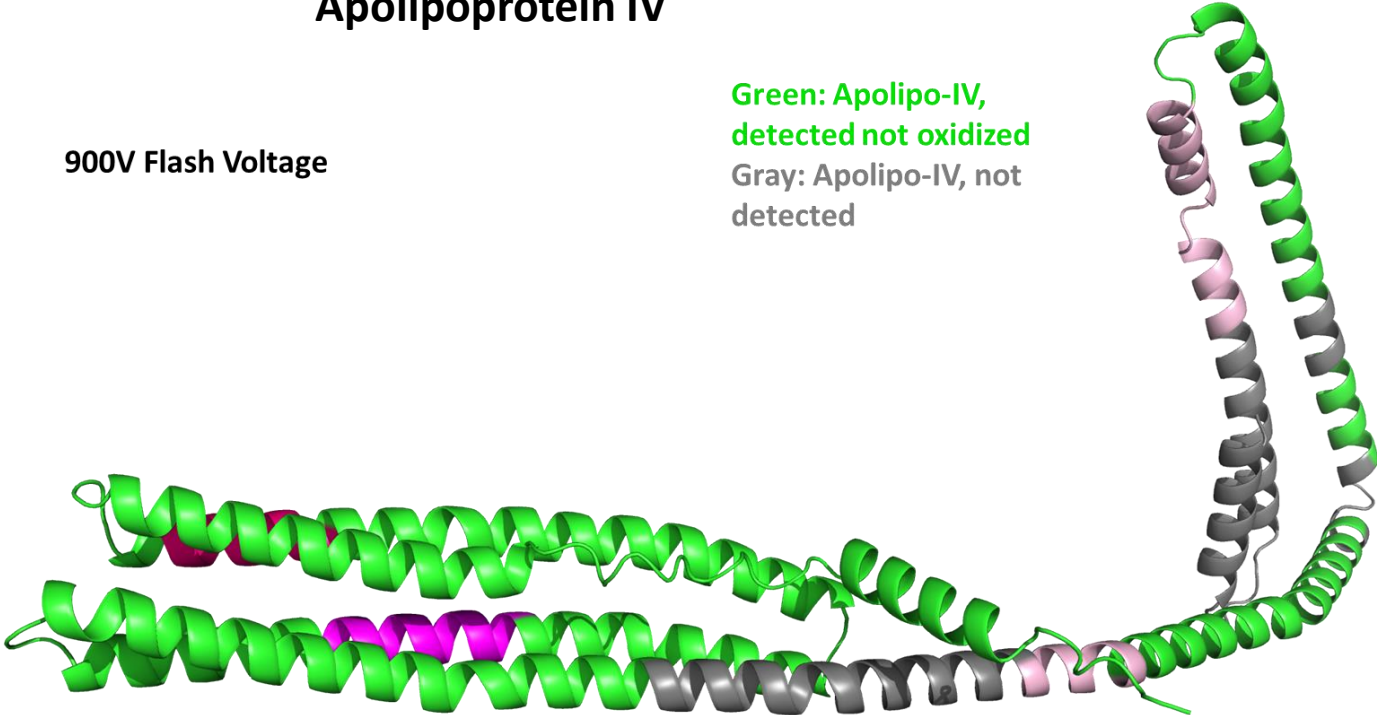
Green: SPI regions  
detected not oxidized



# Apolipoprotein IV

900V Flash Voltage

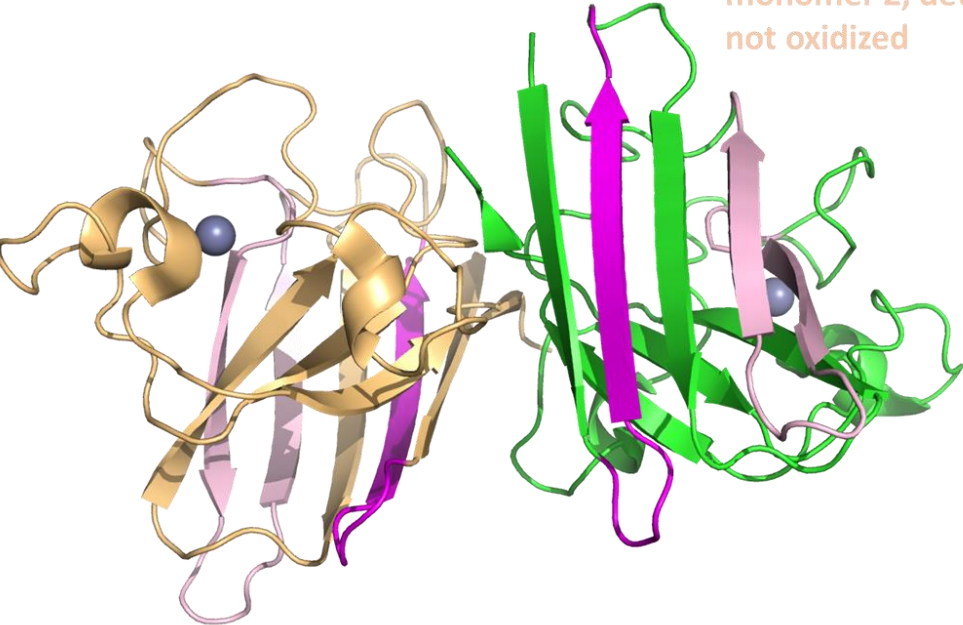
Green: Apolipo-IV,  
detected not oxidized  
Gray: Apolipo-IV, not  
detected



# Superoxide Dismutase

900V Flash Voltage

Green: SOD monomer  
1, detected not oxidized  
Lt. Orange: SOD  
monomer 2, detected  
not oxidized



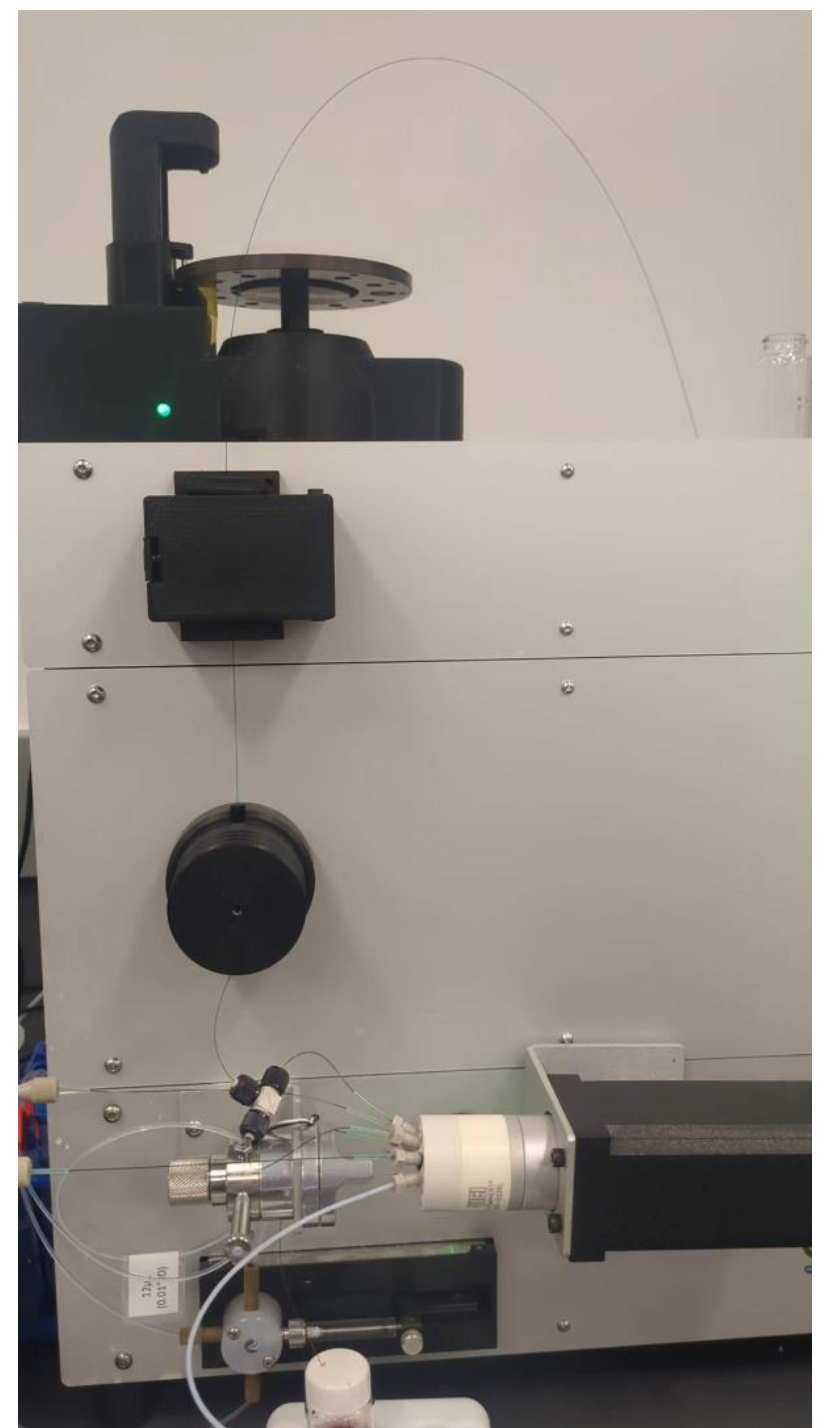
Avg. Oxidations per Peptide





# Current Work

- Automated fluidics handling
  - Custom software fluidics control
  - Automatic low-volume switching valve
  - Hardware integration with Fox system
- Dosimetry methods
- Proteomic complexity and dynamic range
- Workflows for targeted analysis from blood



# Acknowledgements

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## Funding



NIGMS

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The logo for the Jones Research Group, featuring the word "JONES" in large white letters with a yellow and orange wave passing through the letter "O". Below it is a stylized bar chart with a vertical line and an asterisk, and the text "Research Group" in white.

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Lyle Tobin



Aaron Sharp



Prof. Lisa M. Jones



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