

### Regulatory Perspectives on Structural Characterization of Gene Therapy Products

Andrew Byrnes, Ph.D. Director, Division of Gene Therapy 1 Office of Gene Therapy Office of Therapeutic Products, FDA CBER

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### **Overview of Cell and Gene Therapy Products**

Full and empty adeno-associated virus (AAV) capsids

**Deamidation of AAV capsid proteins** 

# Diversity of products regulated by CBER's Office of Therapeutic Products



#### Gene therapies (GT)

Ex vivo genetically modified cells

Non-viral vectors (e.g., plasmids)

Replication-deficient viral vectors (e.g., adenovirus, adeno-associated virus, lentivirus)

Replication-competent viral vectors (e.g., measles, adenovirus, vaccinia)

Microbial vectors (e.g., Listeria, Salmonella)

#### Stem cells/stem cell-derived

Adult (e.g., hematopoietic, neural, cardiac, adipose, mesenchymal) Perinatal (e.g., placental, umbilical cord blood)

Fetal (e.g., neural)

Embryonic

Induced pluripotent stem cells (iPSCs)

#### **Products for xenotransplantation**

**Functionally mature/differentiated cells** (e.g., retinal pigment epithelial cells, pancreatic islets, chondrocytes, keratinocytes)

#### **Combination products**

Engineered tissues/organs

#### Therapeutic vaccines and other antigen-specific active immunotherapies

#### **Blood- and Plasma-derived products**

Coagulation factors Fibrin sealants Fibrinogen Thrombin Plasminogen Immune globulins Anti-toxins Venom antisera for scorpions, snakes, and spiders

Devices

Tissues

# FDA

### Cell and gene therapies: new INDs per year Excluding expanded access



# FDA-approved cellular and gene therapy products

#### Full list:

https://www.fda.gov/vaccines-blood-biologics/cellular-gene-therapy-products/approved-cellular-and-gene-therapy-products

#### **Examples:**

Luxturna – AAV vector
Imlygic – oncolytic HSV
Adstiladrin – adenovirus vector
<b>Vyjuvec</b> – HSV vector
Kymriah – CAR T cell
Zynteglo – hematopoietic stem cell gene therapy
Casgevy – genome-edited hematopoietic stem cell gene therapy
Hemacord – hematopoietic progenitor cells from cord blood
Stratagraft – allogeneic cultured keratinocytes and fibroblasts in bovine collagen
Lantidra – allogeneic pancreatic islets
Amtagvi – tumor-derived autologous T cell immunotherapy

# Adeno-associated virus (AAV) vectors

#### **Protein capsid**

- 25 nm diameter protein shell, 3.8 MDa
- Just 3 proteins, no lipids
- A variety of post-translational modifications
  - Deamidation, phosphorylation, N-terminal truncation, others

#### **DNA** genome

- Single-stranded DNA genome, up to 4.7 kB
  - For very short genes, AAV vectors can be designed to form a half-length "self-complementary" dsDNA genome
  - For long genes, there are some clever ways to split into 2 or 3 vectors
- All viral genes are deleted and replaced with a transgene cassette AAV vectors cannot replicate
- Inefficient genome packaging into capsids
  - High percentage of empty AAV capsids, with no DNA inside
  - Some capsids contain truncated genomes, host cell DNA, or plasmid DNA





## **FULL AND EMPTY AAV CAPSIDS**



## The vast majority of AAV capsids are empty – no DNA inside

Most manufacturing methods include some removal of empty capsids

# Potential for empty capsids to interfere with cell-based assays

# AAV capsids (both full and empty) can be toxic in large amounts

Short-term toxicities due to complement activation

Thrombotic microangiopathy, a few days after administration

Medium-term toxicities due to T cell responses against capsid proteins Hepatotoxicity, including liver failure, a few weeks after administration

These toxicities have been very difficult to model in animals And unpredictable manifestation in clinical trials

# Which physical properties of capsids can be exploited to improve AAV capsid purity?

**Density** – separate using ultracentrifugation Cesium chloride or iodixanol density gradients Extremely efficient separation Can remove empty capsids almost completely

Challenging to automate and scale up



**Charge** – separate using anion exchange chromatography DNA-containing capsids have slightly different surface charge Chromatography is a familiar technology, and easy to automate But separation is inefficient, especially at large scale Diffusion of AAV vectors is slow, so best to use methods that rely on convective mass transfer Monoliths and membrane absorbers, instead of resins

Separation parameters need to be fine-tuned and tightly controlled for each product Full and empty capsid pl differences are very small – need shallow elution gradients FDA

#### Impurities should be identified, controlled, and ideally removed

# FDA advisory committee meeting in September 2021 to discuss the safety of AAV gene therapies

https://www.fda.gov/advisory-committees/advisory-committee-calendar/cellular-tissue-and-gene-therapies-advisory-committee-september-2-3-2021-meeting-announcement

#### Currently, there is no regulatory limit for empty capsids

Different products have vast differences in dose, route of administration, capsid serotype properties Many AAV products have substantial amounts of empty capsids, without undue toxicity Not possible to predict safe levels of empty capsids before starting clinical studies

#### Therefore, regulatory focus is on characterizing and controlling empty capsids

Different lots of a product should have consistent impurity profiles

Empty capsids

Non-vector DNA inside capsids (host cell DNA, plasmid DNA)

Lot release assays should have adequate performance to quantify impurities

Manufacturing processes should be designed to consistently clear impurities

After manufacturing process changes, the product should retain a similar (or better) impurity profile Scale up, changes to cells, chromatography changes, etc.

# Analytical methods for empty capsids

- Transmission electron microscopy
- Cryo-electron microscopy
- A260/A280 absorbance
- Ratio of capsids (e.g., ELISA) to genomes (e.g., qPCR)
- Anion exchange chromatography
- Analytical ultracentrifugation
- Charge-detection mass spectrometry
- Mass photometry

These techniques can

 distinguish among full, empty, and partially-full capsids

# Partially-full capsids



#### AUC is the most widely-used technique for resolving partially-full capsids

#### **Typical AAV vector with ssDNA genome** full empty 92S Normalized C(S) 0.8 62S 0.6 0.4 IF (interferometry) 0.2 0 50 100 150 0

Sedimentation Coefficient



#### Analytical Ultracentrifugation as an Approach to Characterize Recombinant Adeno-Associated Viral Vectors

Brenda Burnham, Shelley Nass, Elton Kong, MaryEllen Mattingly, Denise Woodcock, Antonius Song, Samuel Wadsworth,<sup>†</sup> Seng H. Cheng, Abraham Scaria, and Catherine R. O'Riordan<sup>\*</sup>

Gene Therapy, Genzyme, a Sanofi Company, Framingham, Massachusetts.

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Typical self-complementary AAV vector with dsDNA genome







### **AAV vectors are heterogeneous**

Most capsids are empty

Some capsids contain full vector genomes

Self-complementary AAV vectors can have substantial percentages of partially-full capsid

Some capsids contain host cell DNA or plasmid DNA

### Capsids that do not contain full genomes are impurities

When possible, use purification methods that reduce empty capsids

Quantitate and control capsids forms and DNA impurities in capsids



# **DEAMIDATION OF AAV CAPSID PROTEINS**

# Protein deamidation and AAV vectors Decreased potency





Amide group of an asparagine (or a glutamine) side chain is lost after nucleophilic attack from an adjacent main-chain amide

The intermediate succinimidyl undergoes hydrolysis

Results in amino acid change from asparagine to aspartic acid or isoaspartic acid (1:3)

Or less frequently: from glutamine to glutamic acid or pyroglutamic acid

More likely when Asparagine is followed by Glycine (NG sites)



Giles et al. (2018) *Deamidation of Amino Acids on the Surface of Adeno-Associated Virus Capsids Leads to Charge Heterogeneity and Altered Vector Function.* Mol. Ther. 26:2848

# Ronit Mazor laboratory at FDA CBER





What impact does deamidation have on the immunogenicity of AAV vectors?

FDA

# Immune responses to AAV vectors



# Protein deamidation and AAV vectors Altered human T cell response to capsid



**Mazor lab:** Bing et al. (2022) *Differential T cell immune responses to deamidated adeno-associated virus vector.* Mol. Ther. Meth. Clin. Devel. 24:255

# Protein deamidation and AAV vectors Altered human T cell response to capsid



#### Altered T cell response after substituting Asn with Asp / iso-Asp in an AAV9 capsid peptide



This particular donor's T cells show:

- Higher response with aspartate in peptide
- Reduced response with isoaspartate



**Mazor lab:** Bing et al. (2022) Differential T cell immune responses to deamidated adeno-associated virus vector. Mol. Ther. Meth. Clin. Devel. 24:255

# Protein deamidation and AAV vectors

Deamidation increases T cell responses in some donors, decreases in others



**Mazor lab:** Bing et al. (2022) *Differential T cell immune responses to deamidated adeno-associated virus vector.* Mol. Ther. Meth. Clin. Devel. 24:255



# Protein deamidation and AAV vectors

### High level of deamidation in AAV vectors

More deamidation over time during storage Residue-dependent variability

### **Deamidation may decrease AAV vector activity**

Deamidation may increase or decrease the immunogenicity of AAV vectors

T cell assays that do not include deamidated peptides may underestimate T cell responses to vectors





### **AAV vectors have complex heterogeneity**

- Full, empty, and partial capsids
  - Empty capsids have no activity, need to be controlled
- Post-translational modifications
  - The impact of PTMs on AAV vector quality is not completely understood PTMs may affect vector activity and immunogenicity

### We recommend:

Develop manufacturing processes that yield vectors with consistent impurity profiles

Develop assays to quantitate and control capsid forms

Characterize post-translational modifications and seek to understand their impact on product quality

# **Contact Information**

Andrew Byrnes

Andrew.Byrnes@fda.hhs.gov

• Regulatory Questions:

OTP Main Line – 240 402 0685 Email: OTPRPMS@fda.hhs.gov

• OTP (OTAT) Learn Webinar Series:

http://www.fda.gov/BiologicsBloodVaccines/NewsEvents/ucm232821.htm

- CBER website: <u>www.fda.gov/BiologicsBloodVaccines/default.htm</u>
- **Phone:** 1-800-835-4709 or 240-402-8010
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