

Computational Methods for Comparison of NMR Spectra

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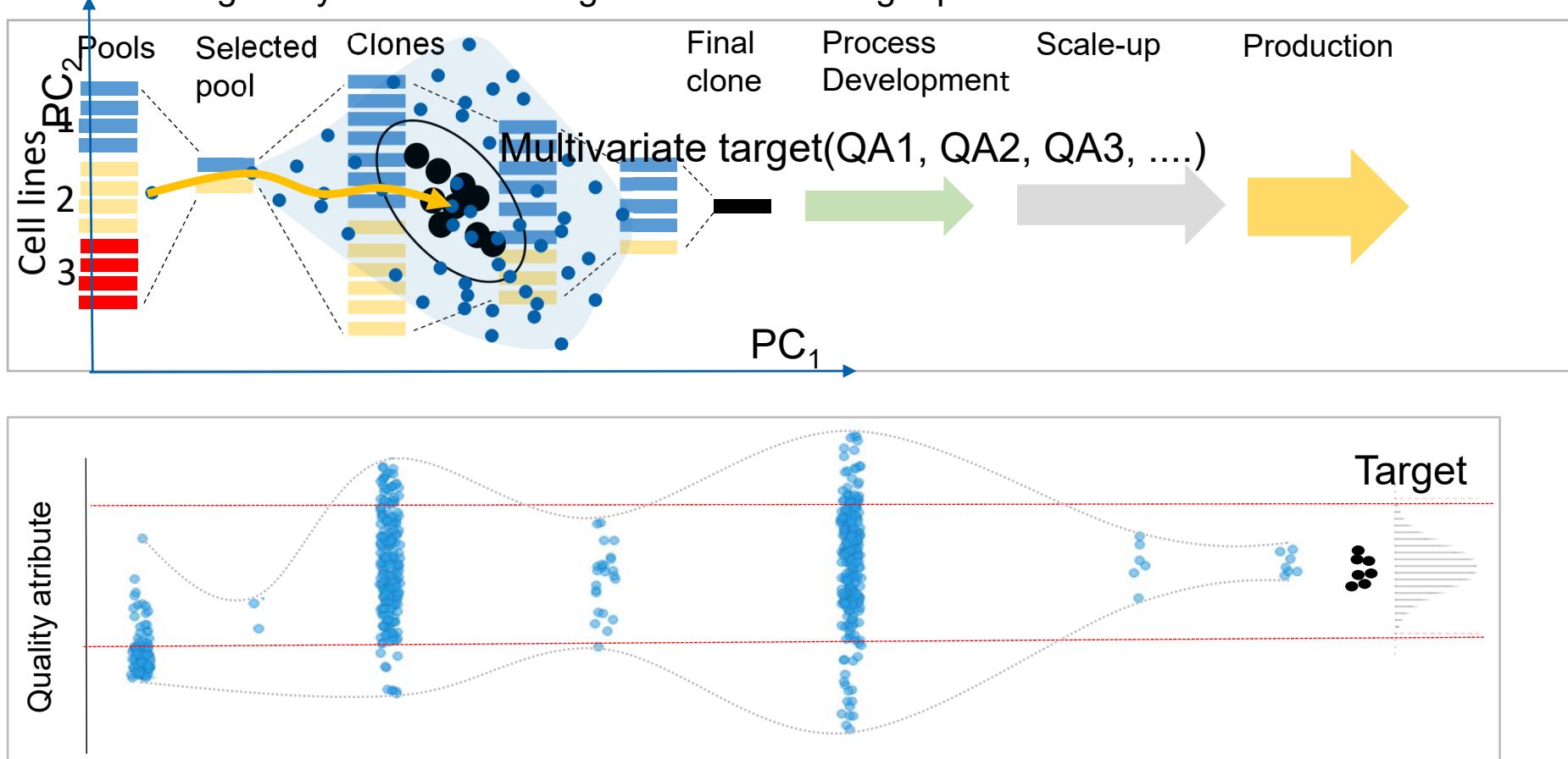


Agenda

- Introduction
- NMR fingerprinting experiments
- Computational methods for spectral comparisson
- High throughput 1D NMR workflow for stability screening
- Conclusion

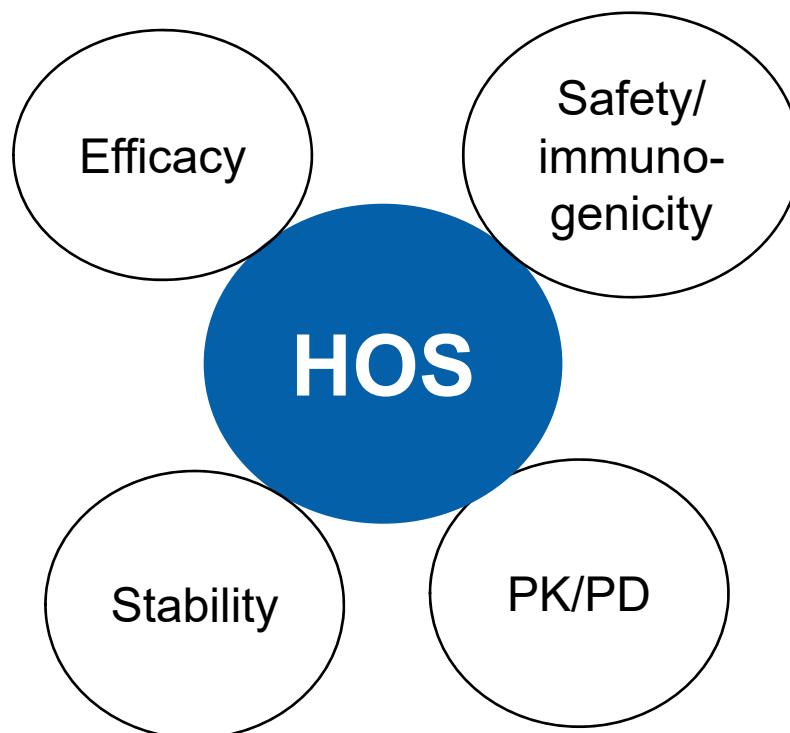
Introduction

Biological drugs are produced using a complex bioprocess which results in structural heterogeneity. Biosimilars target multivariate target profile.



QA = f(raw materials, process parameters, formulation, conditions,...)

Higher order structure



Biological drugs	
Size	Large (mixture of related molecules)
	High molecular weight
Structure	Complex (heterogeneous), defined by the exact manufacturing process
Modification	Many options
Manufacturing	Produced in living cell culture
	Difficult to control from starting material to final API
	Impossible to ensure identical copy
Characterisation	Very difficult to completely characterize due to molecular composition and heterogeneity
Stability	Unstable, sensitive to external conditions
Immunogenicity	Could be immunogenic

Ref: <http://www.gabionline.net/Biosimilars/Research/Small-molecule-versus-biological-drugs>; Declerck PJ. GaBI J. 2012;1(1)

Analytical characterization used during development of biosimilars

- X-ray crystallography
- Nuclear magnetic resonance (NMR)
- Cryoelectron microscopy
- Circular dichroism
- Fourier-transform infrared spectroscopy
- Intact MS
- Mass spectrometry
- Peptide mapping, ...
- Hydrogen/deuterium exchange with mass spectrometry
- Dynamic light scattering (DLS)
- Microcalorimetry
- CEX, CIEF acidic/basic variants
- Biolayer interferometry
- Electron tomography
- LC glycation
- Analytical ultracentrifugation
- Peptide mapping
- Field-flow fractionation, mutations, glycation
- Free-electron laser scattering
- SEC/FFF/AUC aggregation
- Raman spectroscopy
- Size-exclusion chromatography
- Target binding
- Static light scattering
- Cytotoxicity (ADCC, ADCP, CDC)
- UV/fluorescence spectroscopy

• voltage electron microscopy

• Metal-enhanced fluorescence (MEF) or

chemiluminescence (MEC)

Secondary/Tert./Quart. structure:

• Anilinonaphthalene sulfonate (ANS) binding

• Chromatography using interactive resins

• Single-cell sensing

• X-Ray crystallography

• CD

• Single-molecule fluorescence spectroscopy, ...

• FT-IR,...

• H-D exchange,

Total number available

Yearly available

Primary structure:

Intact MS

Peptide mapping, ...

Hydrogen/deuterium exchange with mass spectrometry

Dynamic light scattering (DLS)

Microcalorimetry

CEX, CIEF acidic/basic variants

Biolayer interferometry

Electron tomography

LC glycation

Analytical ultracentrifugation

Peptide mapping

Field-flow fractionation, mutations, glycation

Free-electron laser scattering

SEC/FFF/AUC aggregation

Raman spectroscopy

Size-exclusion chromatography

Target binding

Static light scattering

Cytotoxicity (ADCC, ADCP, CDC)

UV/fluorescence spectroscopy

Bioactivity (potency):

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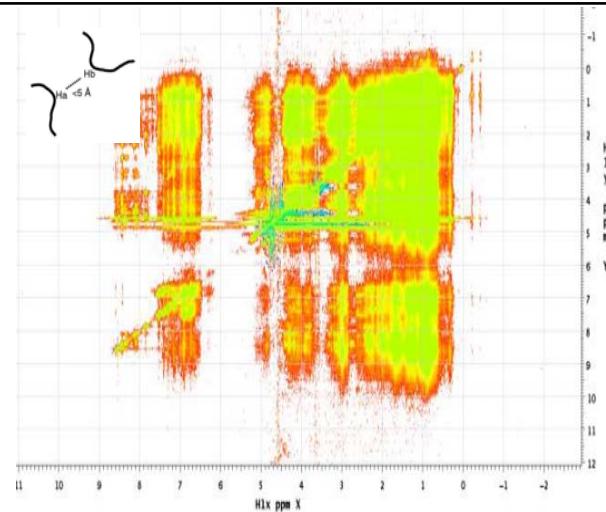
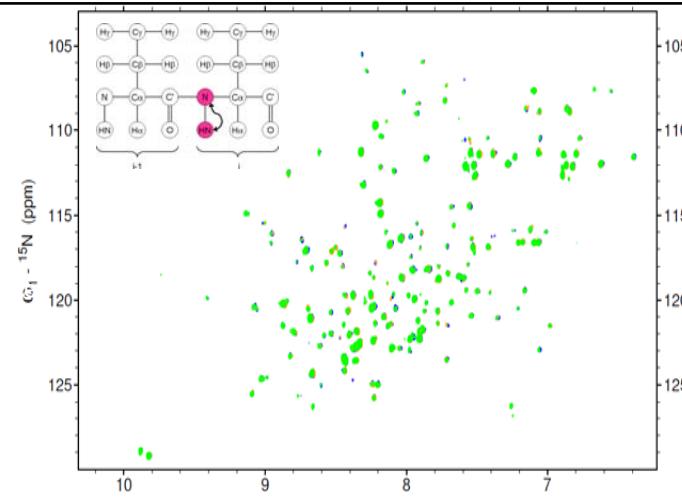
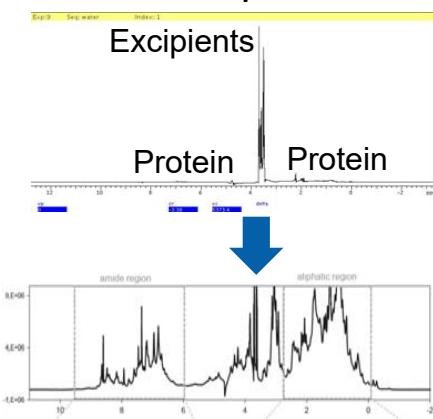
Size-exclusion chromatography

Target binding

NMR fingerprinting experiments

NMR experiment	Advantages	Disadvantages
^1H spectra	Fast, simple	Overlapping signals, non-selective towards excipients
^1H - ^1H NOESY	Higher resolution than 1D experiments, through space dipolar interaction	Overlapping signals, non-selective towards excipients, complex analysis
^1H - ^{15}N gsHSQC (US/NUS)	Smaller number of signals / better dispersion than NOESY	Low sensitivity for non-labeled samples (0.37% nat. occurring ^{15}N isotope)
^1H - ^{13}C gsHSQC (US/NUS)	Smaller number of signals / better dispersion than NOESY, works well for large proteins	Low sensitivity for non-labeled samples (1.11% nat. occurring ^{13}C isotope), only 6 residues have methyl groups
^1H - ^{13}C -(sf)HMQC	Smaller number of signals / better dispersion than NOESY, works well for large proteins	Low sensitivity for non-labeled samples

^1H NMR spectrum



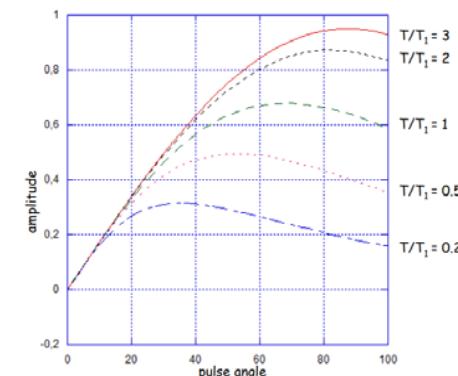
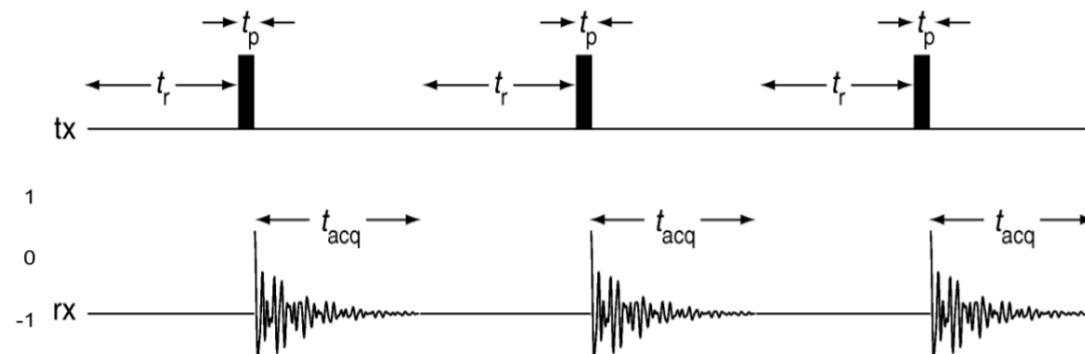
Bodenhausen, G. et al. Chem. Phys. Letters. 69 (1): 185–189 (1980)
<http://www.cryst.bbk.ac.uk/PPS2/projects/schirra/html/2dnmr.htm>
Keeler, J. (2010). Understanding NMR Spectroscopy (2nd ed.). Wiley. pp. 184–187.
<https://www.chemie.uni-hamburg.de/nmr/insensitive/tutorial/en.lproj/>

*Abbreviations:
US- uniform sampling
NUS- non-uniform sampling
sf-SOFAST

NOVARTIS

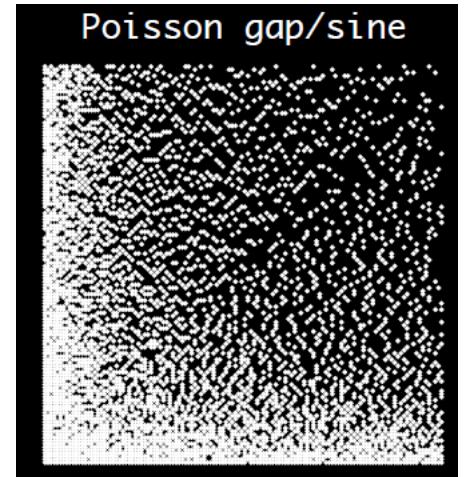
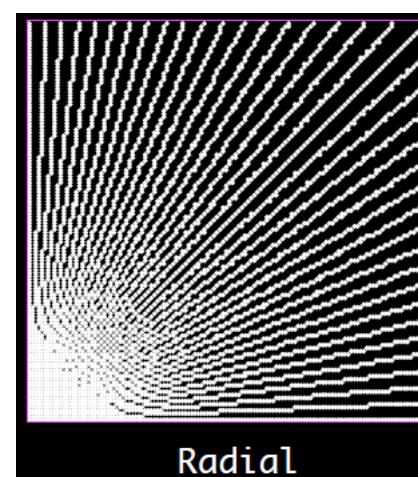
Tricks to increase S/N:

- Temperature (45-50°C)
- Optimize concentration
- Digestion
- CH₃ relaxation
 - 3-fold symmetry
 - Fast rotation around the C-C bond
- Ernst Angle: SOFAST HMQC, BEST-HSQC



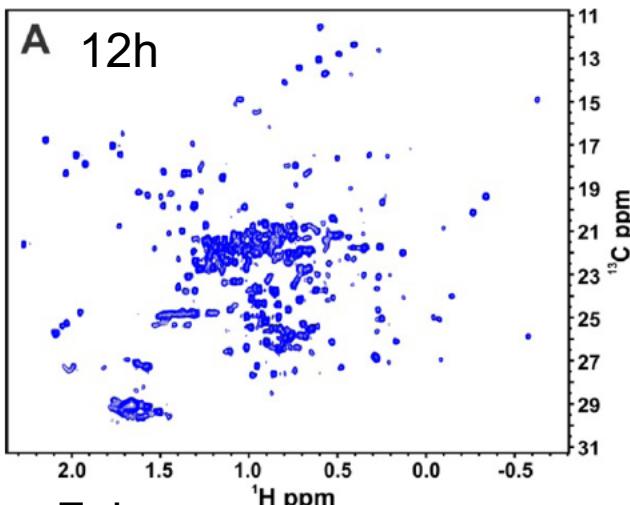
• non-uniform sampling

partial sampling of points in the indirect dimension(s) (triangular, spiral, Poisson Gap, and Burst sampling)

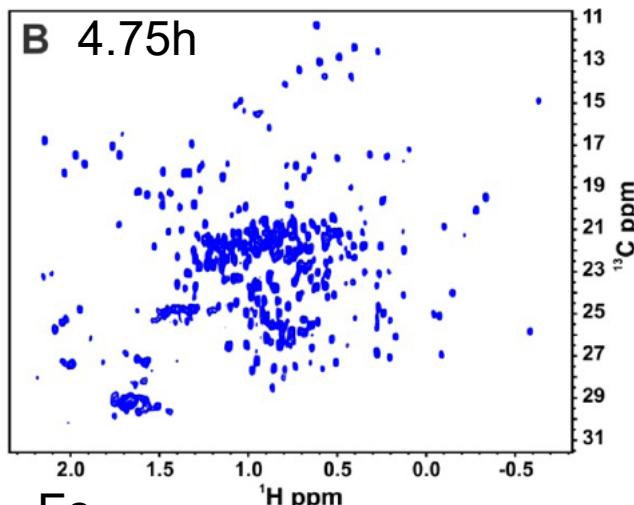


NMR methyl fingerprint method of an intact mAb and fragments at natural isotopic abundance

Intact NISTmAb

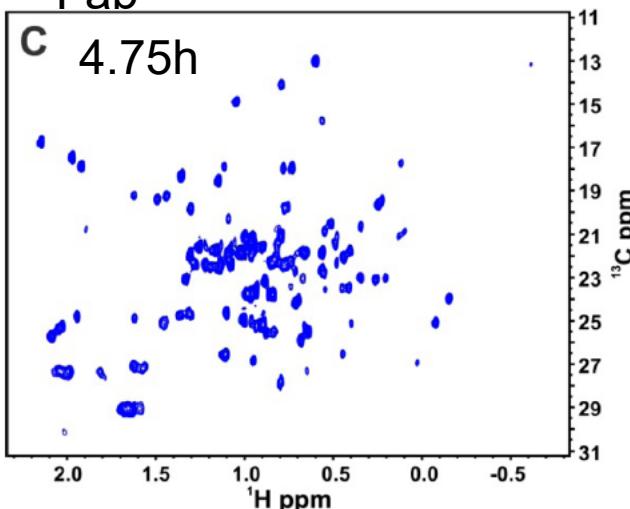


2Fab + Fc

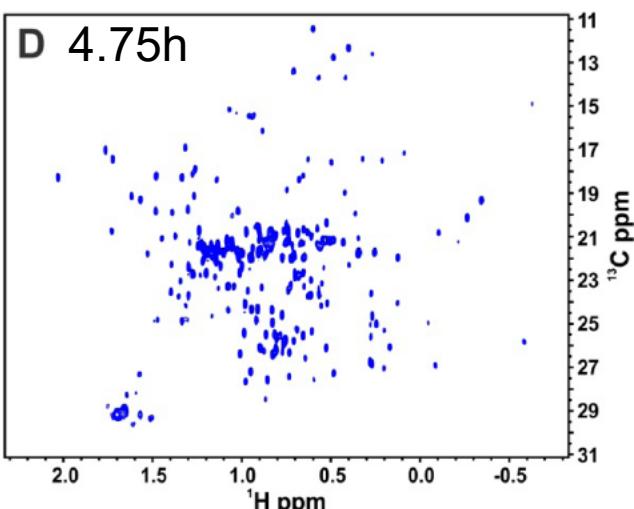


NIST,
900 MHz, 50oC
100% coverage of
intact mAb in 12h

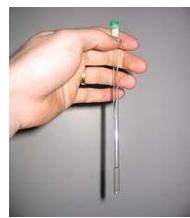
Fab



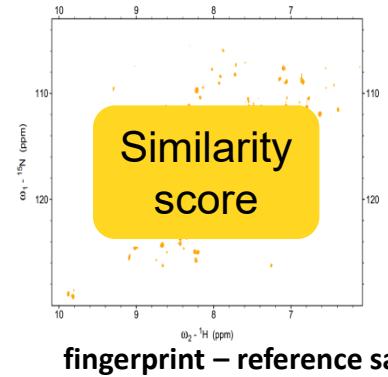
Fc



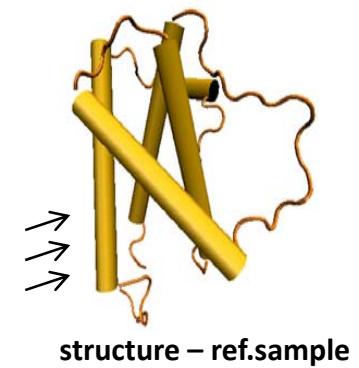
NMR comparability



Reference sample
(originator)



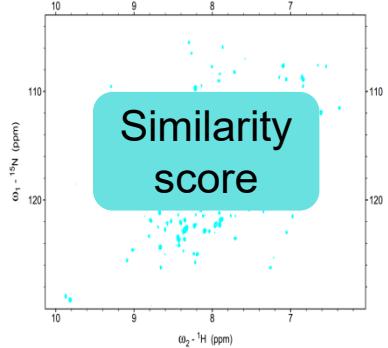
fingerprint – reference sample



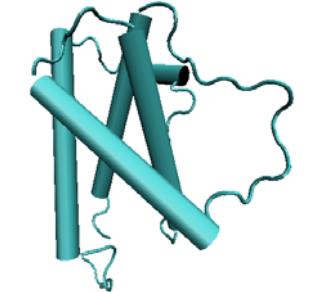
structure – ref.sample



Biosimilar sample



fingerprint – biosimilar sample



structure biosimilar sample

Computational methods for comparison of NMR fingerprints:

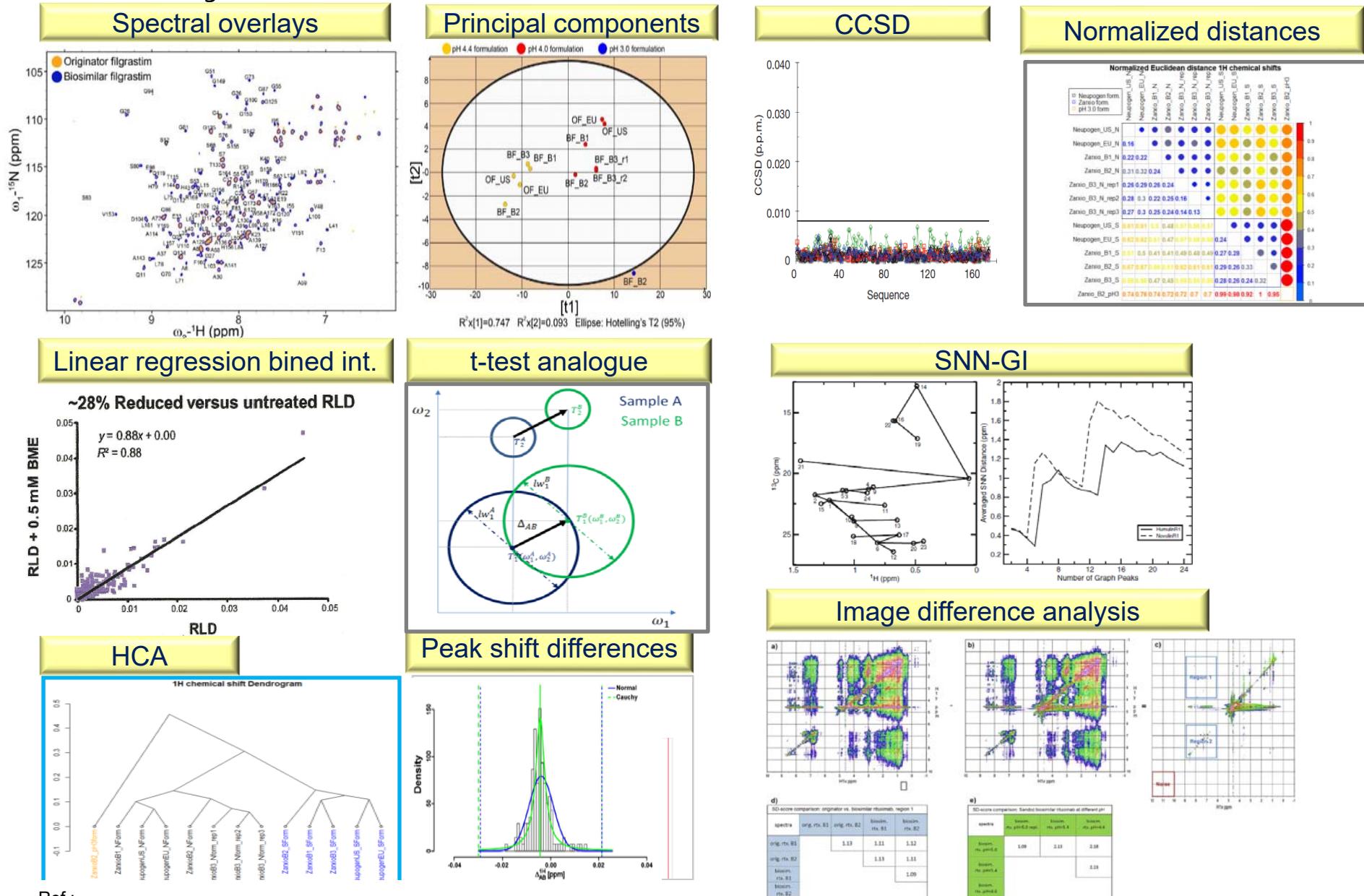
- 1D/amide/methyl fingerprint spectra overlays
- Principal component analysis
- CCSD (combined chemical shift difference)
- t-test analogue for chemical shift quantitation
- Normalized distances
- Peak shift difference- population analysis
- linear regression analysis of binned NMR spectra
- Image analys by spectral subtraction
- Graph invariant sequential nearest neighbours (SNN-GI)
- Tucker3
- Hieararchical clustering

Ref.:

- Japelj, B. et al Sci Rep. 6, 32201 (2016)
Brinson RG, et al. MAbs. 11(1):94-105 (2019)
Ghasriani, H. et al Nat. biotechnol. 34, 139–141 (2016)
Amezcuia, C. Et al. J. Pharm. Sci. 102, 1724–1733 (2013)
Chen, K. Et al. AAPS PharmSciTech, Vol. 19, No. 3, April 2018 (2017)
Župerl, Š et al J. Chem. Inf. Model. 47, 737–743 (2007).
Arbogast, L. Et al Anal Chem 89, 11839-11845 (2017)

- PCA, corr., Image analysis, norm. dist., HCA, t-test an.**
CCSD, PCA
CCSD, PCA
Linear regression on binned NMR spectra – ECHOS-NMR
PCA, GI-SNN, Tucker3
- GI-SNN Noesy spectra**
Linear correlation, PCA

Similarity metrics overview



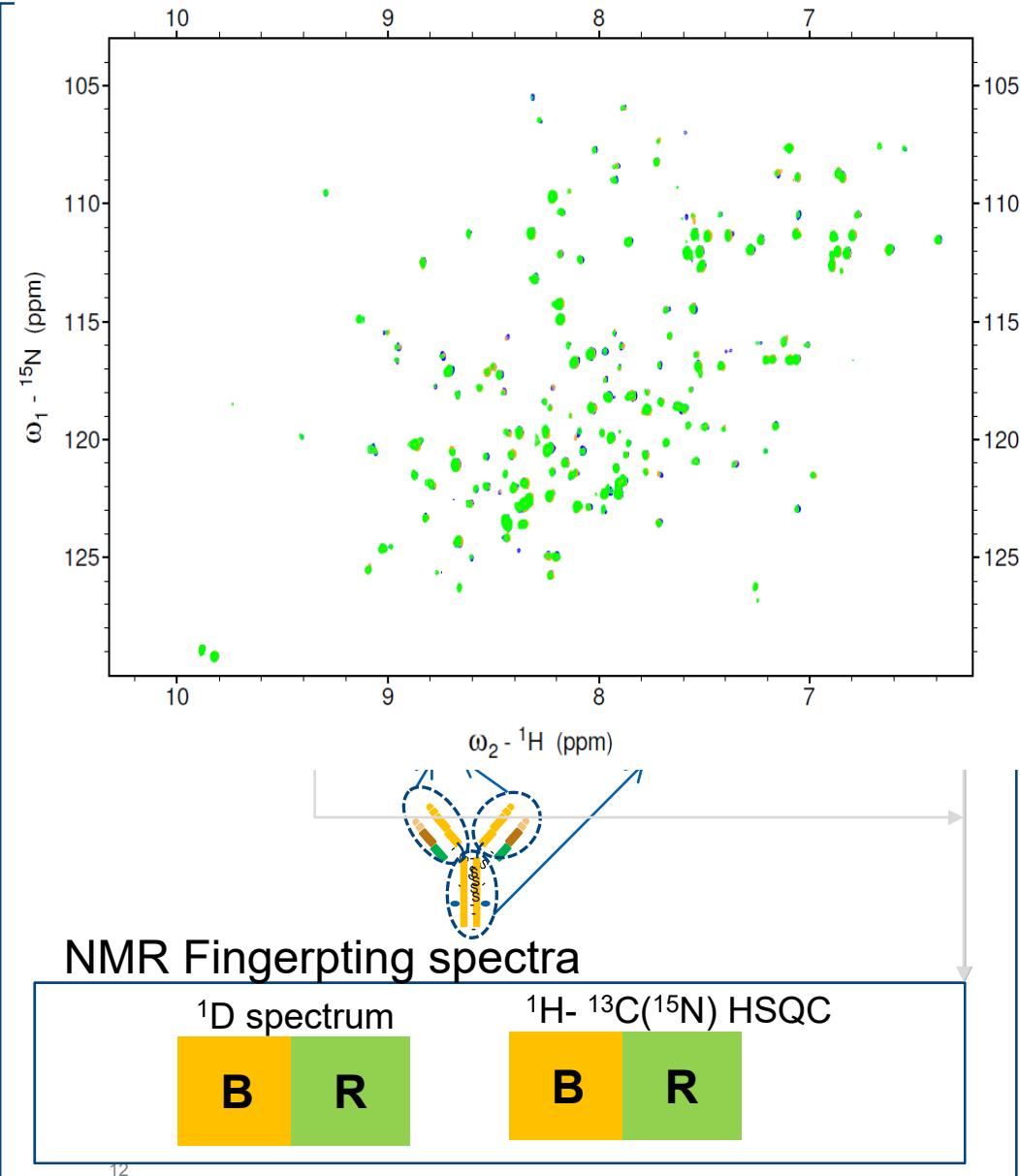
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Japelj, B. et al Sci Rep. 6, 32201 (2016), Amezcua, C. Et al. J. Pharm. Sci. 102, 1724–1733 (2013)

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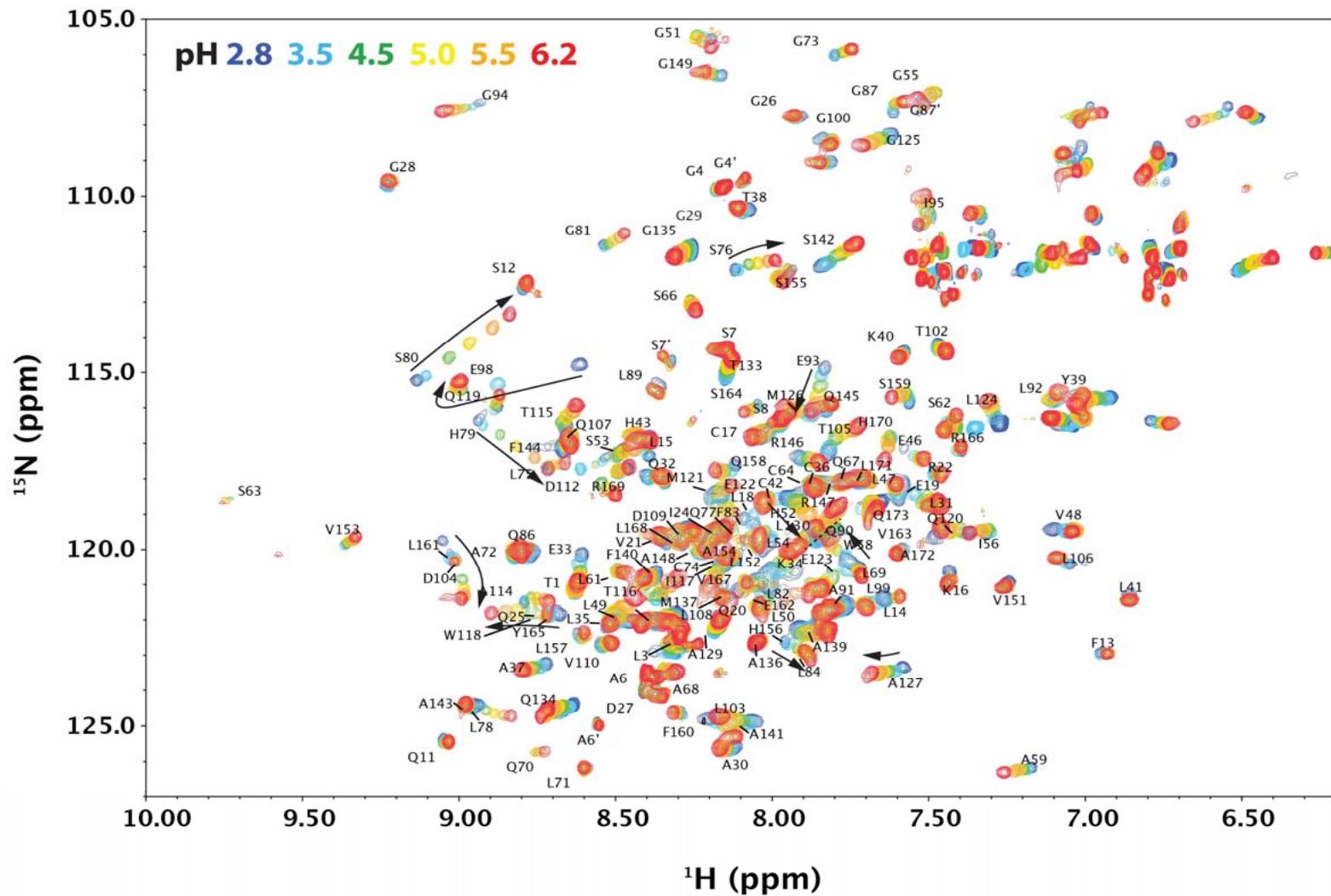
Comparison should be performed in the same environment (buffer)



Legend:

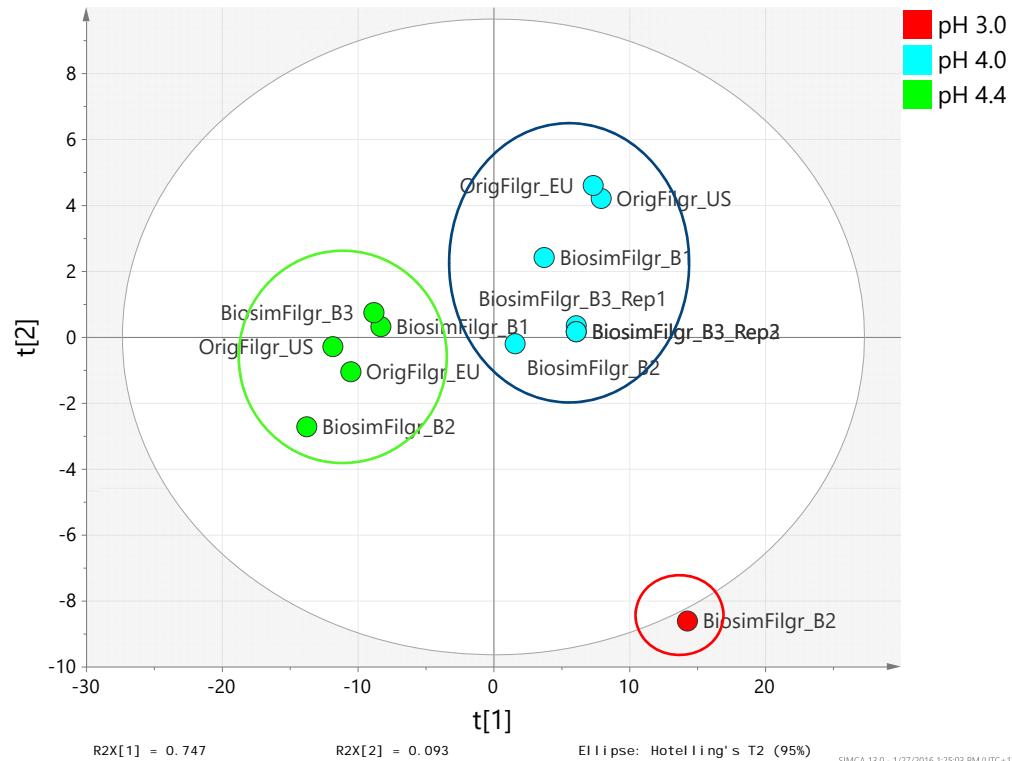
color	Protein	Formulation
Blue	Zarxio(R) Biosimilar filgrastim B1	Originator filgrastim
Yellow	Originator filgr. US	Originator filgrastim
Green	Originator filgr. EU	Originator filgrastim

NMR method sensitivity: pH effect on protein conformation

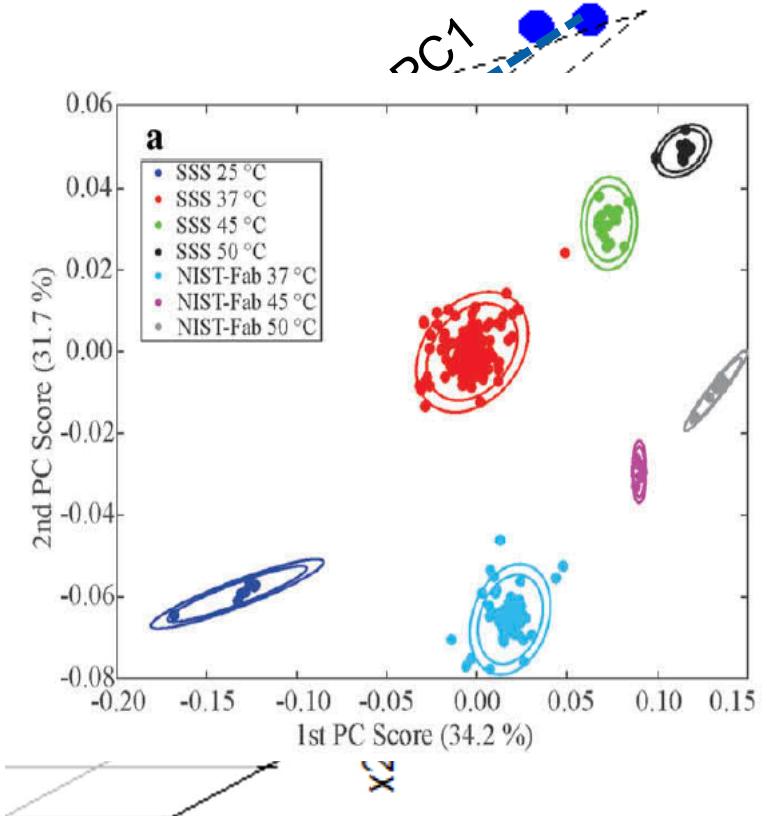


Aubin, Y., Hodgson, D.J., Thach, W.B. et al. *Pharm Res* (2015) 32: 3365, Suppl. Fig4
Japelj, B. et al. *Sci Rep*: 6, 32201 (2016), Suppl. Fig2

Principal component analysis

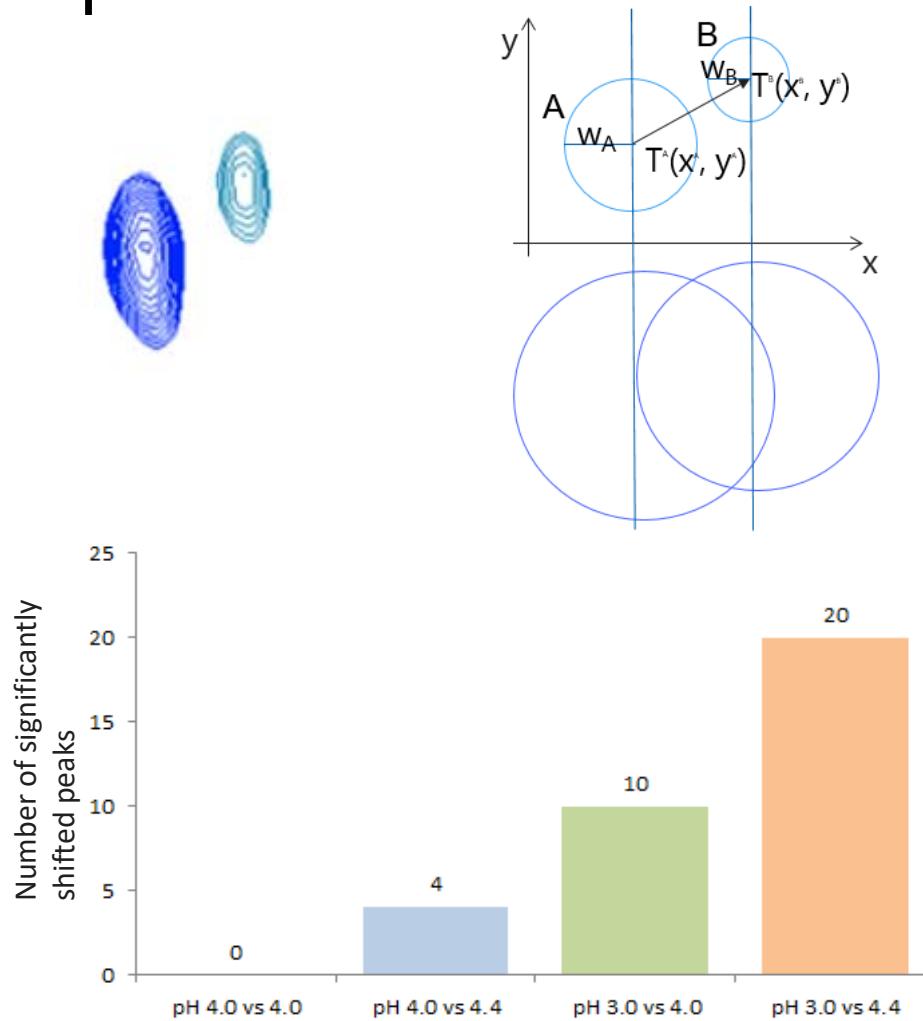


NMR method is sensitive enough to detect small pH induced conformational changes



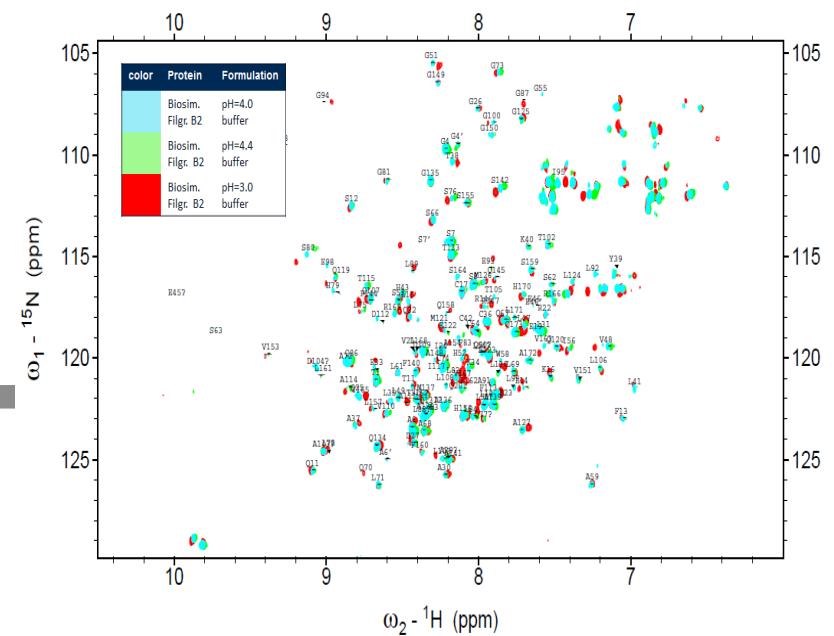
Clustered PCA scatter plots of all peak lists from 354 ^1H - ^{13}C spectra-
26 laboratories
Brinson RG, et al. MAbs. 11(1):94-105
(2019)

t-test analogue for chemical shift quantitation



$$d(T^A, T^B) = \sqrt{(x^A - x^B)^2 + \alpha^2(y^A - y^B)^2}$$

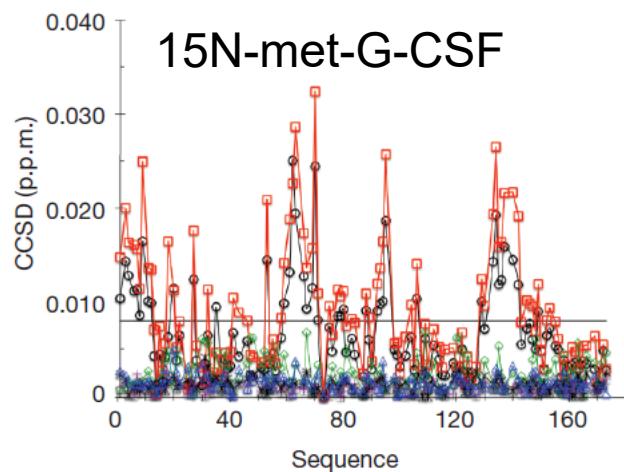
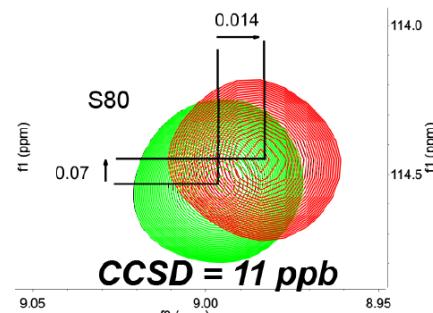
$$t \sim \frac{d(T^A, T^B)}{\sqrt{w_A^2 + w_B^2}}$$



Combined chemical shift difference (CCSD)

$$\text{CCSD} = \sqrt{(0.5 * [(\delta_H)^2 + (\alpha * \delta_N)^2])}$$

$\alpha=0.1$



NIST 900 (plus), NIST 600 (star), HC 700 (circle), HC 600 (square), FDA 500 (diamond) and MPA 600 (triangle)

Temperature calibration

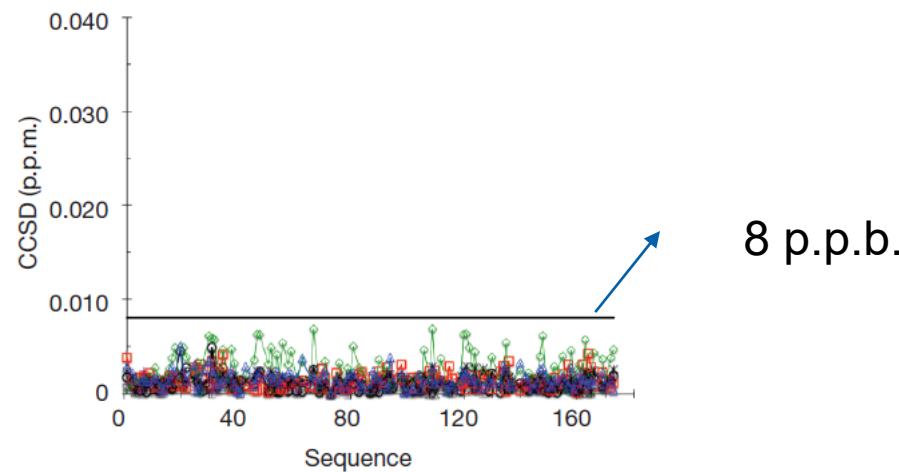
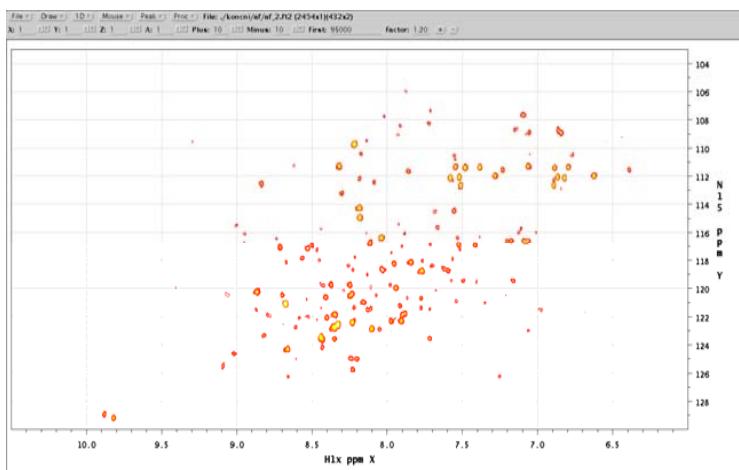


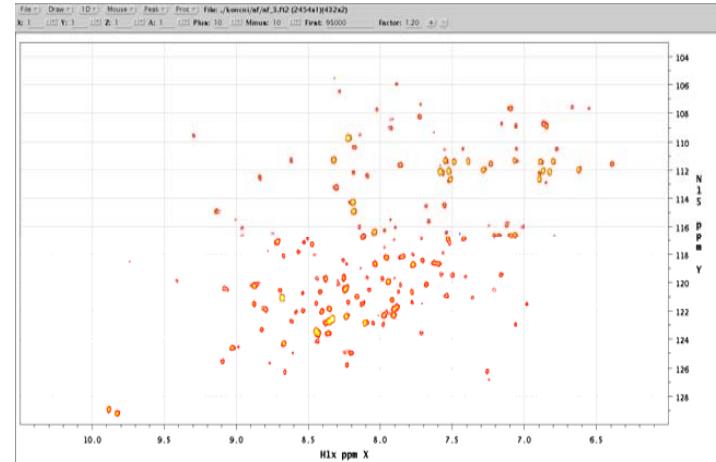
Image difference analysis

Subtraction of G-CSF ^1H - ^{15}N HSQC spectra

Originator filgrastim EU



Biosimilar filgrastim B1



Difference spectrum

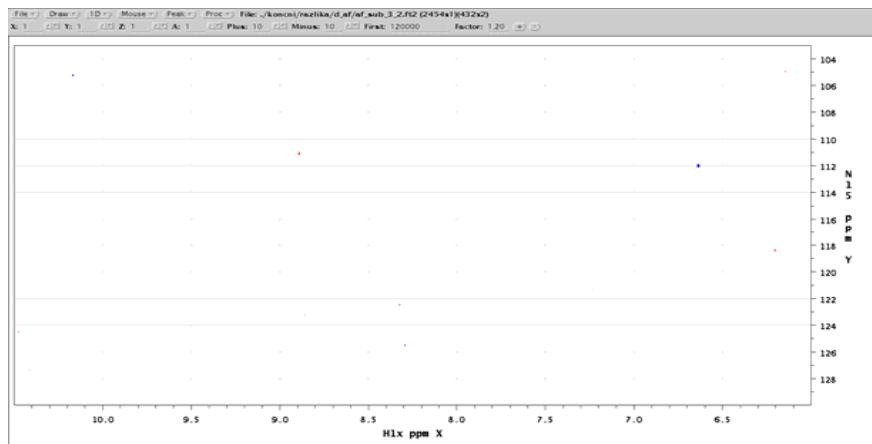
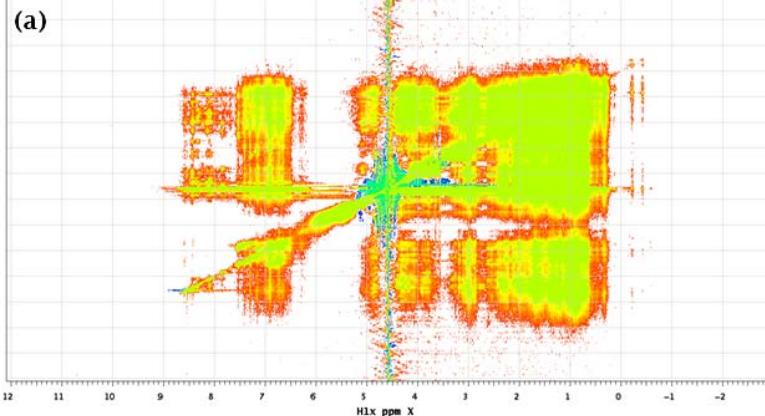


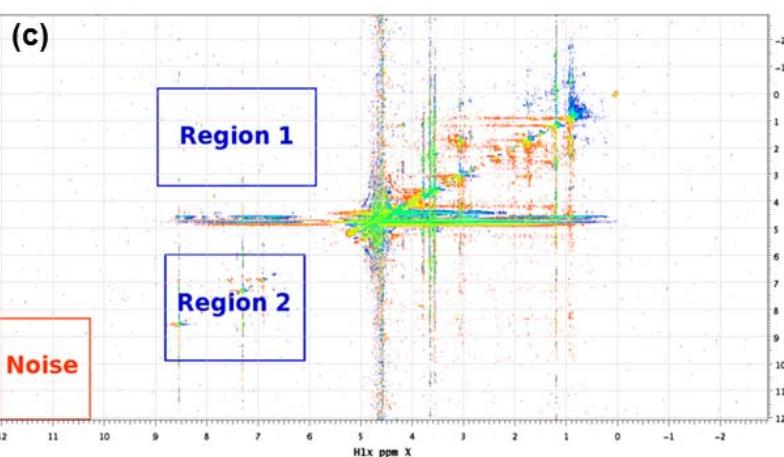
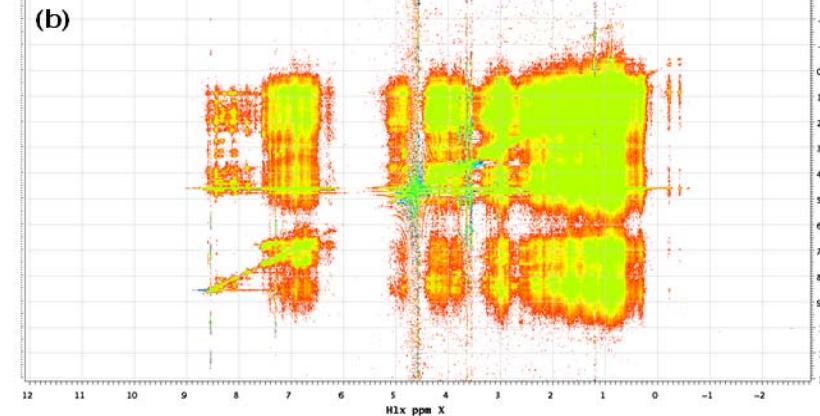
Image difference analysis

Subtraction of rituximab NOESY spectra

NOESY spectrum of full sized Sandoz biosimilar rituximab



NOESY spectrum of full sized originator rituximab



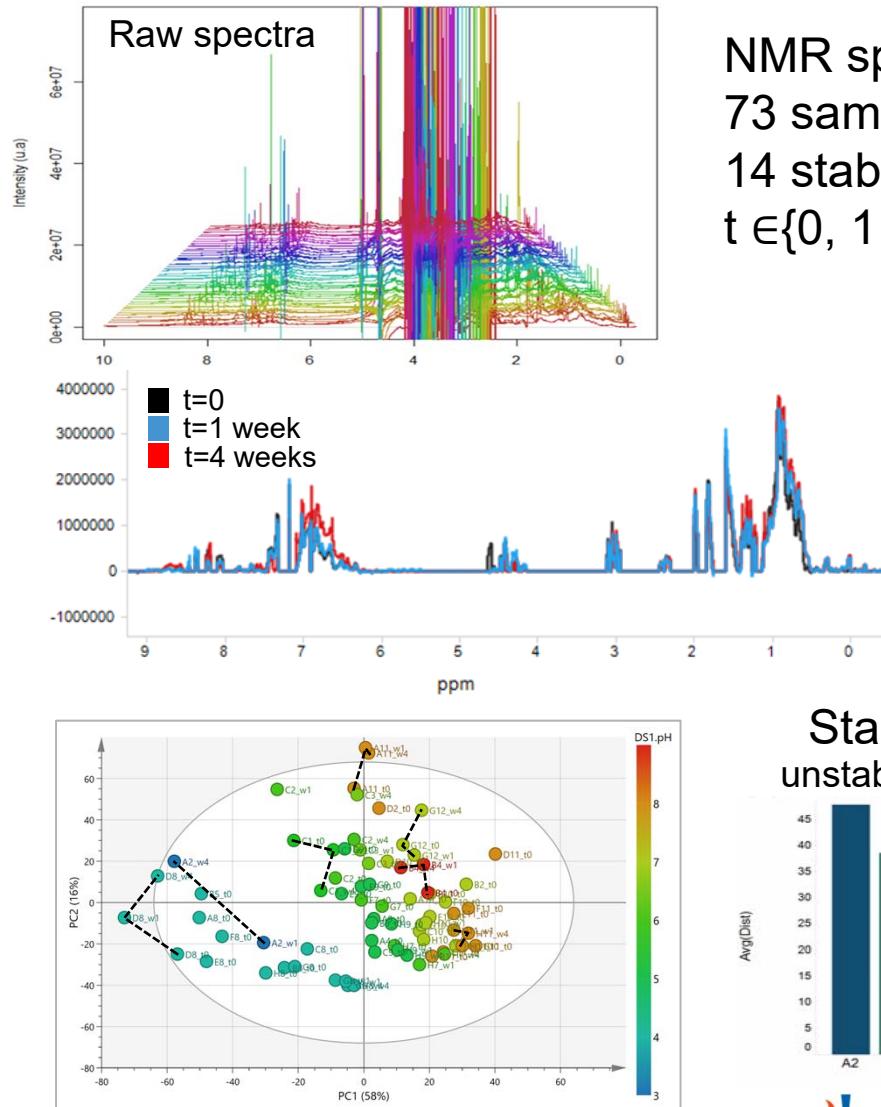
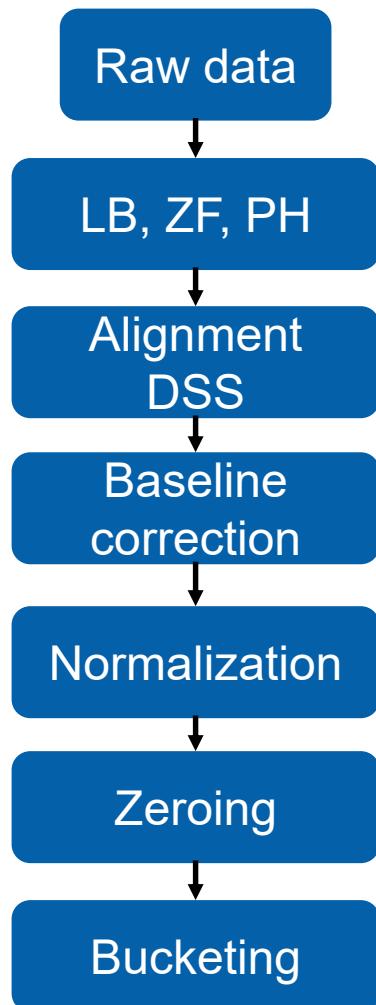
In house software		SBR B1, Orig.Ritux® B1 replicate	SBR B1, Orig.Ritux® B2 pH=5.4	SBR B1, Sandoz biosim. ritux. B1 pH=4.6
SD-scores, region 1 Raw data (FID)				
SBR B1, Orig. Ritux. B1 Fourier transformation	1.09	2.13	2.18	
SBR B1, Orig. Ritux. B2 Automatic phase correction			2.13	
SBR B1, Sandoz biosim. ritux. B1 Automatic baseline correction				

pH change of 0.4 units increases the SD-score
SBR vs Orig. Ritux. SD-score comparable
by ~2-fold
to Orig. Ritux. US vs. EU reference product

$$SD\ score = \sqrt{\frac{signal_{Region\ 1,2}^2}{signal_{Noise}^2}}$$

High throughput 1D NMR workflow for stability screening

Formulation screening: pH, NaCl, Stabilizers, Buffers



NMR spectra,
73 samples
14 stability samples
 $t \in \{0, 1 \text{ week}, 1 \text{ month}\}$



Conclusions

- NMR is a powerful method to compare protein HOS
- can detect conformational changes, changes in formulation, stability changes (PTMs), sequence variants, glycosylation
- methods available for small and large proteins: ^1H - ^{13}C gsHSQC (methyl), ^1H - ^{15}N HSQC (amide) fingerprints + rapid pulsing + NUS
- chemometrics/computational methods available to compare and evaluate differences between NMR spectra for global and peak-to-peak comparison

Acknowledgement

EN-FIST centre of excellence,

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Prof. Janez Plavec



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Technical Research & Development:**

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Drago Kuzman

Jure Senčar

Mitja Zidar

Mateja Salobir

Matej Horvat

Stefan Prasch

Isabel Feuerstein

Johann Holzmann

Prof. Uroš Urleb