

sgRNA sequencing

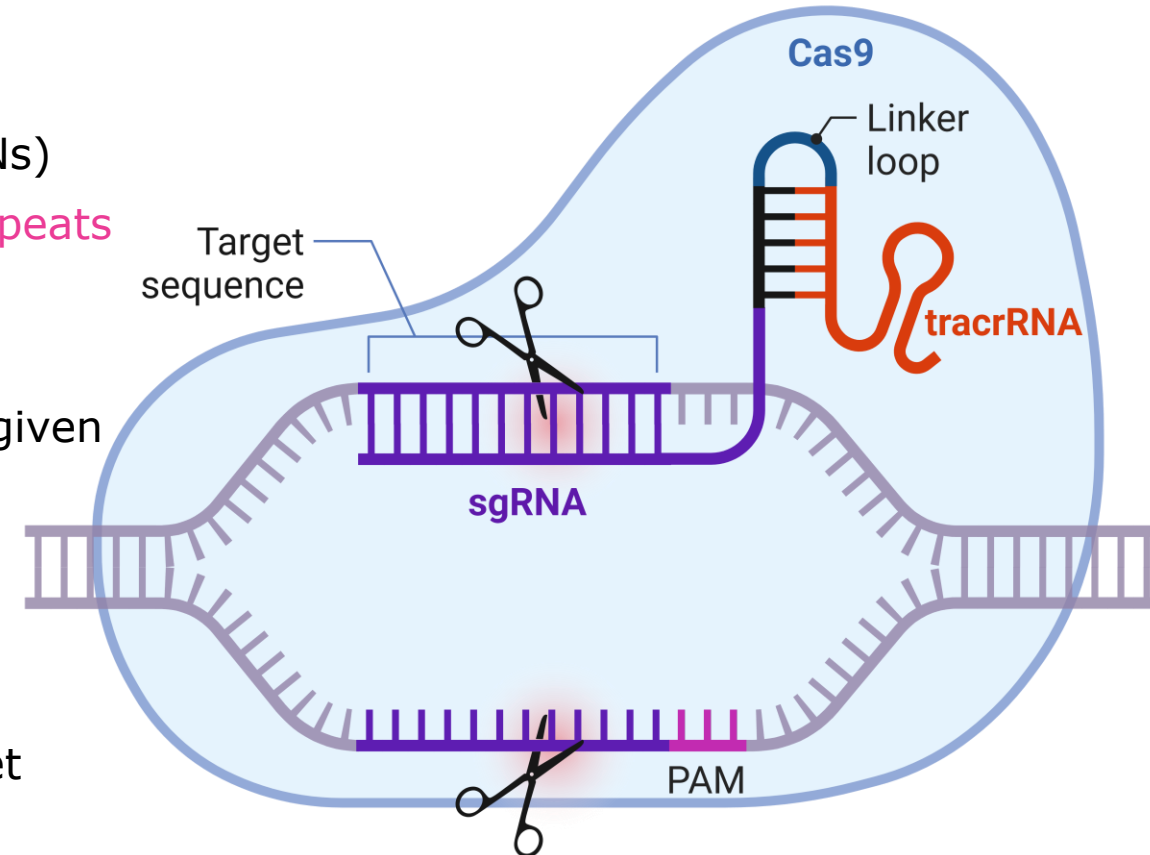
Leveraging an established NGS assay to
sequence single-guide RNAs (sgRNA)

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Cell and Gene Editing

- Current Gene editing technologies:
 - Zinc-finger nucleases
 - Transcription Activator-Like Effector Nucleases (TALENs)
 - Clustered Regularly Interspaced Short Palindromic Repeats (CRISPR) complexes (e.g., CRISPR-Cas9)
- CRISPR has opened the door for gene editing that has given more accessibility to researchers due to its simplicity
- CRISPR-Cas system is composed of:
 - A (CRISPR-associated) Cas nuclease
 - The Cas nuclease is shuttled/guided to intended target sequence by a single-guide RNA (sgRNA)



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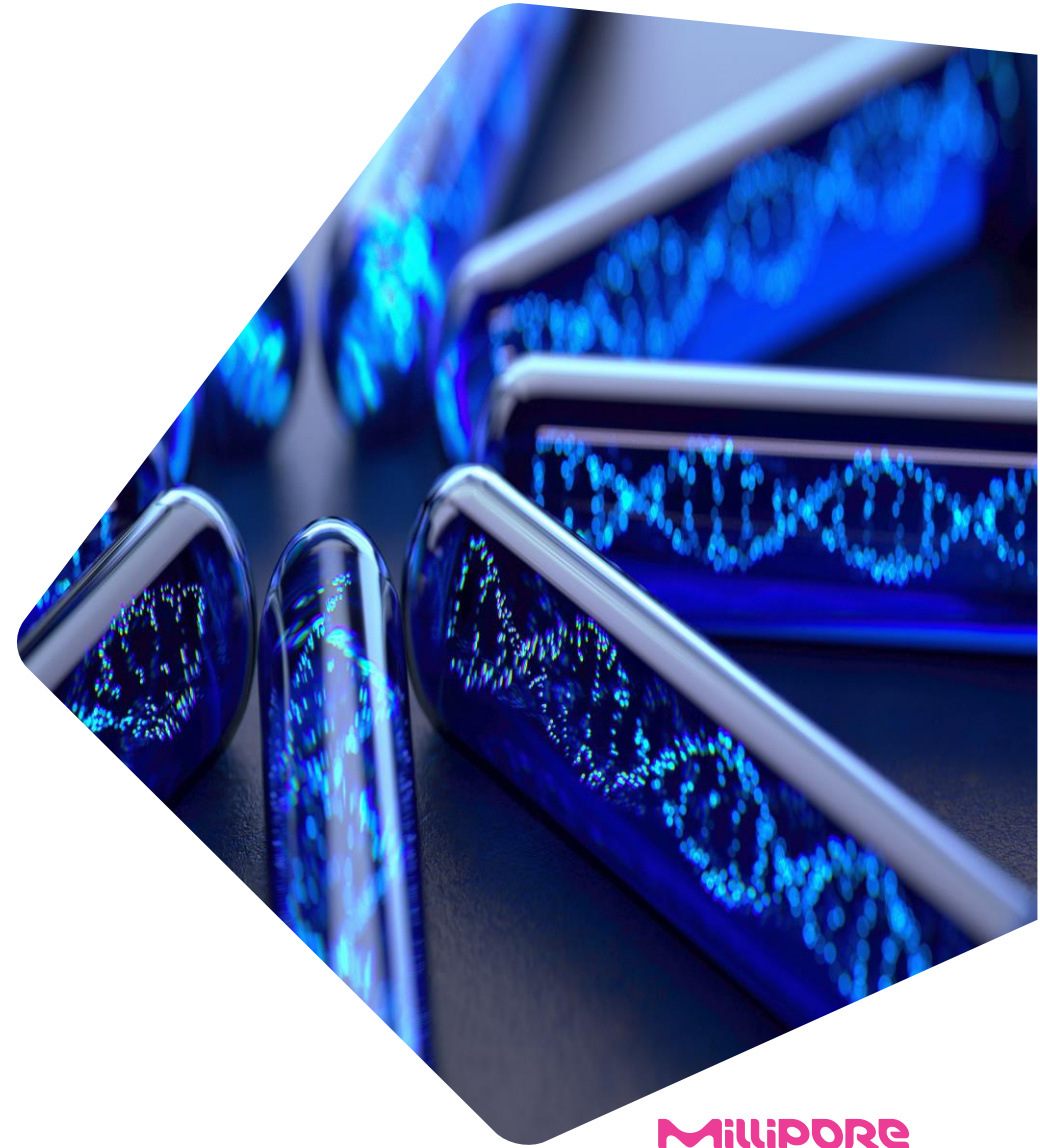
What is a sgRNA?

- The sgRNA is composed of a **CRISPR RNA (crRNA)** and a **trans-activating CRISPR RNA (tracrRNA)** that is compatible with the specific endonuclease
- The customizable part of a sgRNA is the crRNA:
 - 17-24bp sequence complementary to the “**on-target**” site where the dsDNA break and edit should occur
- Unintentional binding of the sgRNA to any other areas → “**off-target**” event
- The design of the sgRNA is a critical step for gene editing experiments

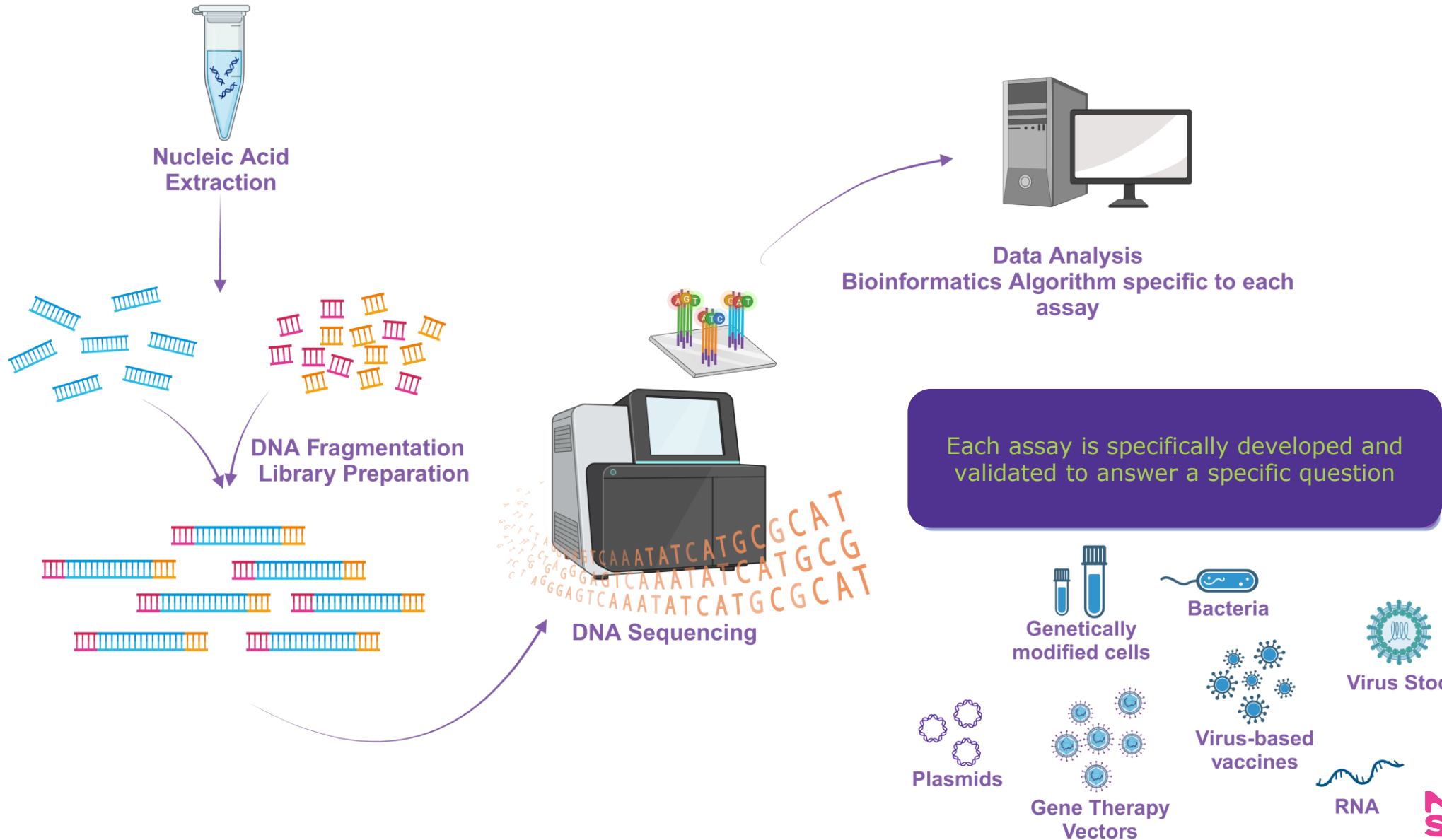


Current QC for sgRNAs and gaps

- Chemical synthesis of sgRNAs can introduce low level contamination → need to measure guide **sequence fidelity** and **purity**
 - Current QC Method:
 - Mass Spec technology
 - Lacks specificity
 - Can miss subtle variations that could contribute to off-target events
- No longer sufficient for regulatory agencies!
- Need ultrasensitive techniques to assess quality, safety and efficacy
 - What about Sanger sequencing?
 - Low sensitivity for variant detection, not high-throughput
 - **NGS**
 - Identify variants down to a 1% presence in the population



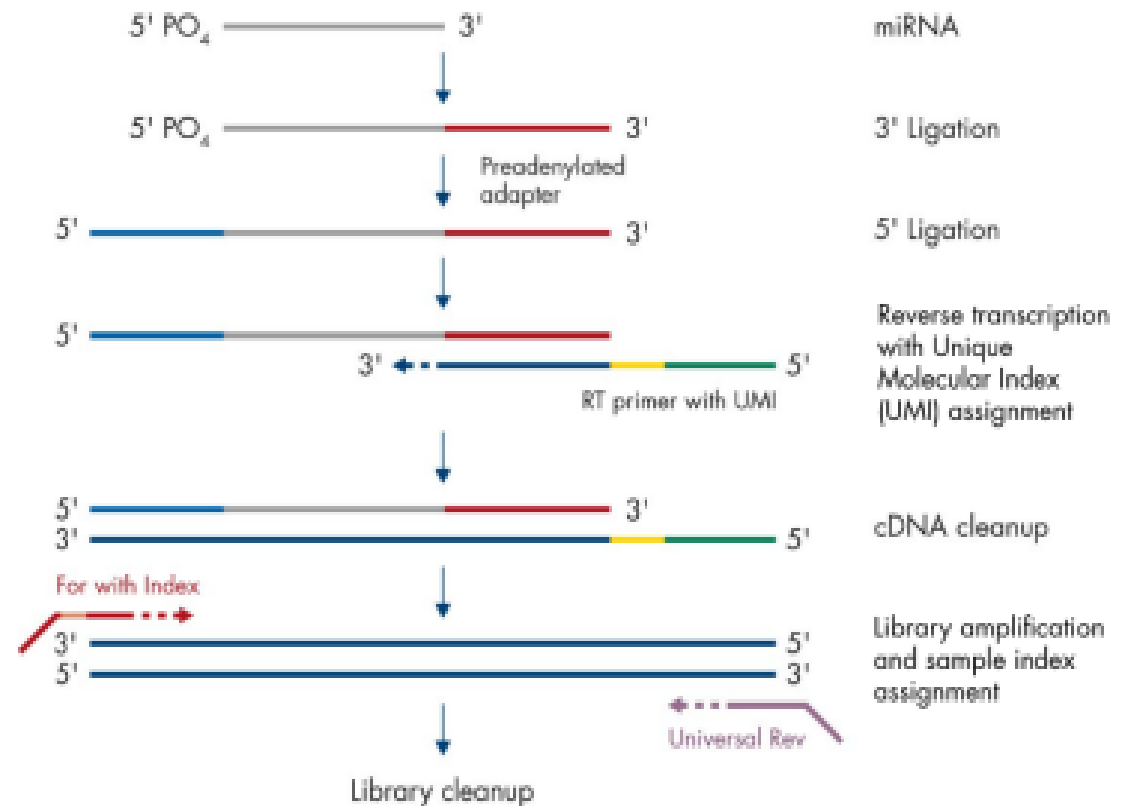
Existing assay



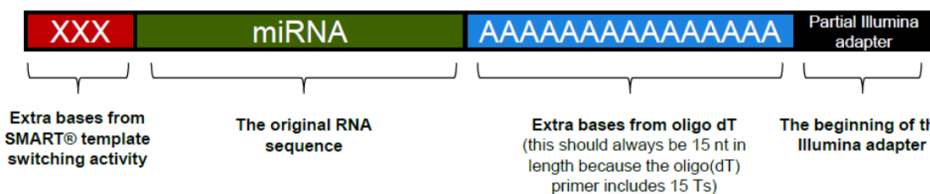
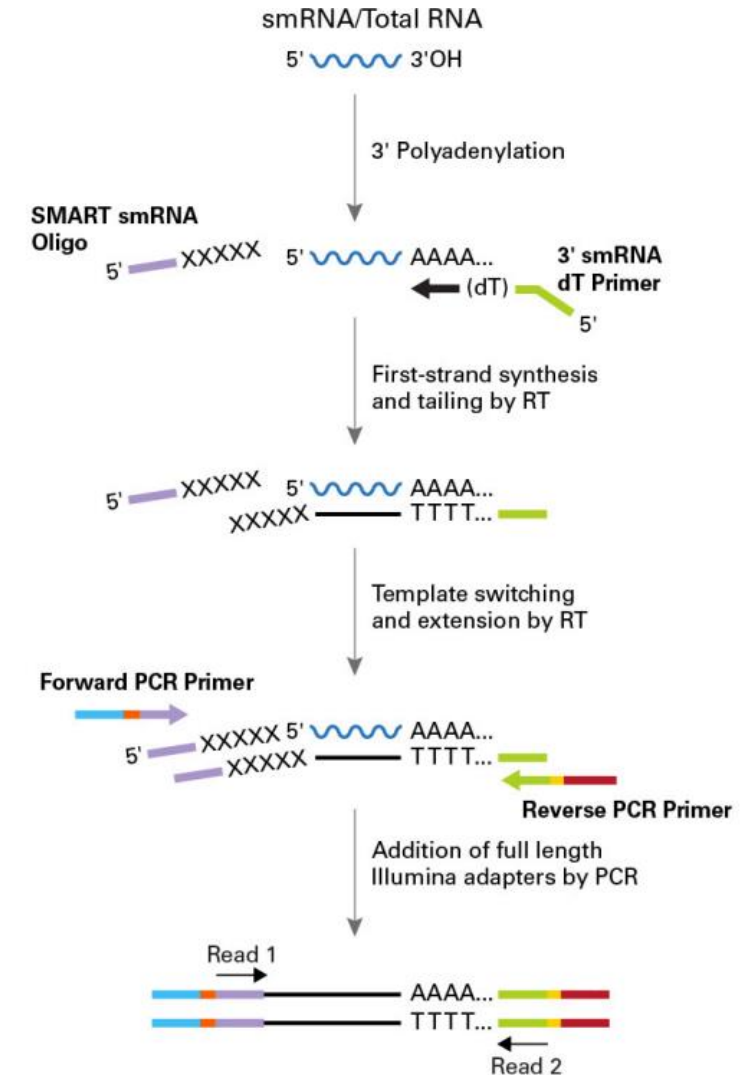
- Identify a kit to generate a compatible library for these short RNA sequences
- Two leading contenders for library preparation kits
- Tested various sgRNA with different modifications to assess and optimize these kits to fit into our current methodology
 - Many sgRNAs have termini modifications to decrease degradation and improve gene editing efficiency
 - Successful libraries can be sequenced using the Illumina Sequencers
- Bioinformatics analysis
 - Sequence Identity, and Variant calling
 - Customization of existing workflow used for identity testing assays

Kit A

- Modified Protocol to capture larger RNAs (~100nt)
- Sequential Ligation Methodology
- Unique Molecular Indexes (UMIs)
- No gel purification
- Possible limitations:
 - Requires 3' hydroxyl and 5' phosphate for ligation



- Methodology:
 - Ligation free
 - 3' Polyadenylation
 - 1st strand synthesis followed by template-switching and extension by RT
- Possible limitations - unambiguous determination of RNA termini not possible due to:
 - poly A tail addition
 - at low frequencies, template switching can add more than 3 nt to cDNA 5' ends and make identifying the beginning of the sequence with 100% confidence not possible



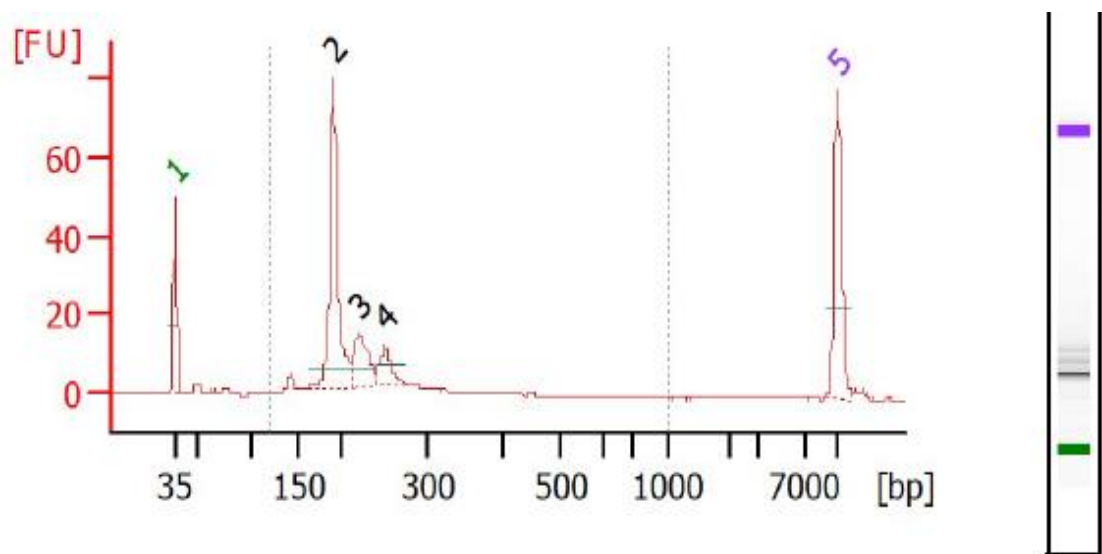
Figures: Takara Bio Inc.

sgRNAs that have been tested

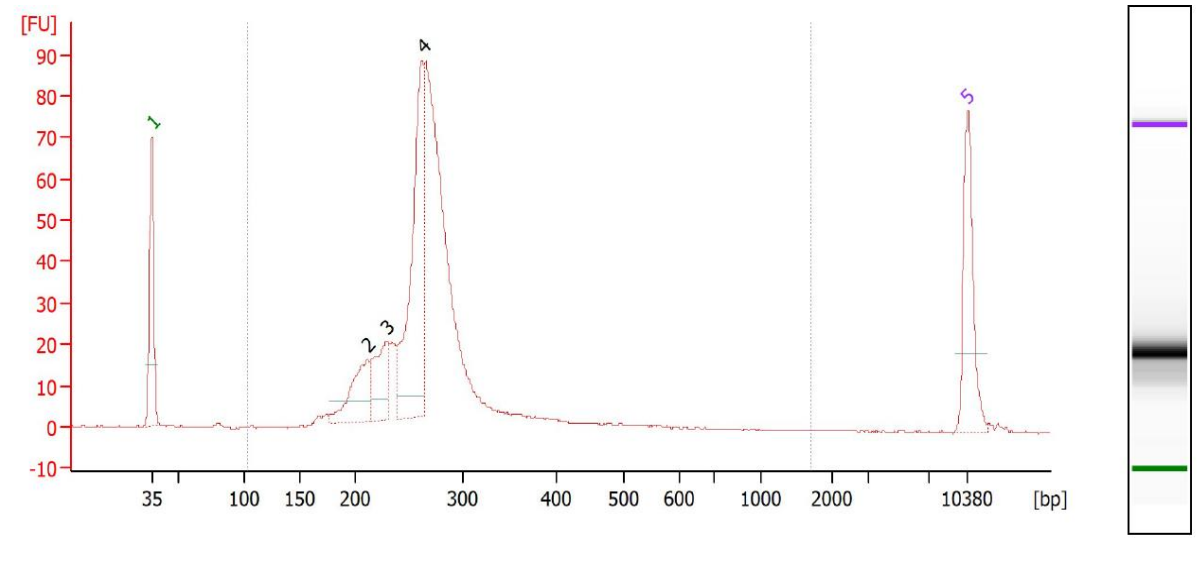
- Purchased sgRNAs from 3 different manufacturers
 - All had similar end modifications:
 - Phosphorothioate bonds on 3' and 5' ends (terminal nucleotides)
 - Protects ends of molecules from degradation → increases editing efficiency
 - 2'Ome
 - Protects from hydrolysis
- Two Client drivers to aid in development
 - Company 1 tested 3 different samples
 - Company 2 tested 8 different samples of varying lengths and modifications

Results – success!

- CNAT was able to successfully generate libraries with both kits



- **Kit A**
- Dilution 1:20
- Average Size: 216 bp

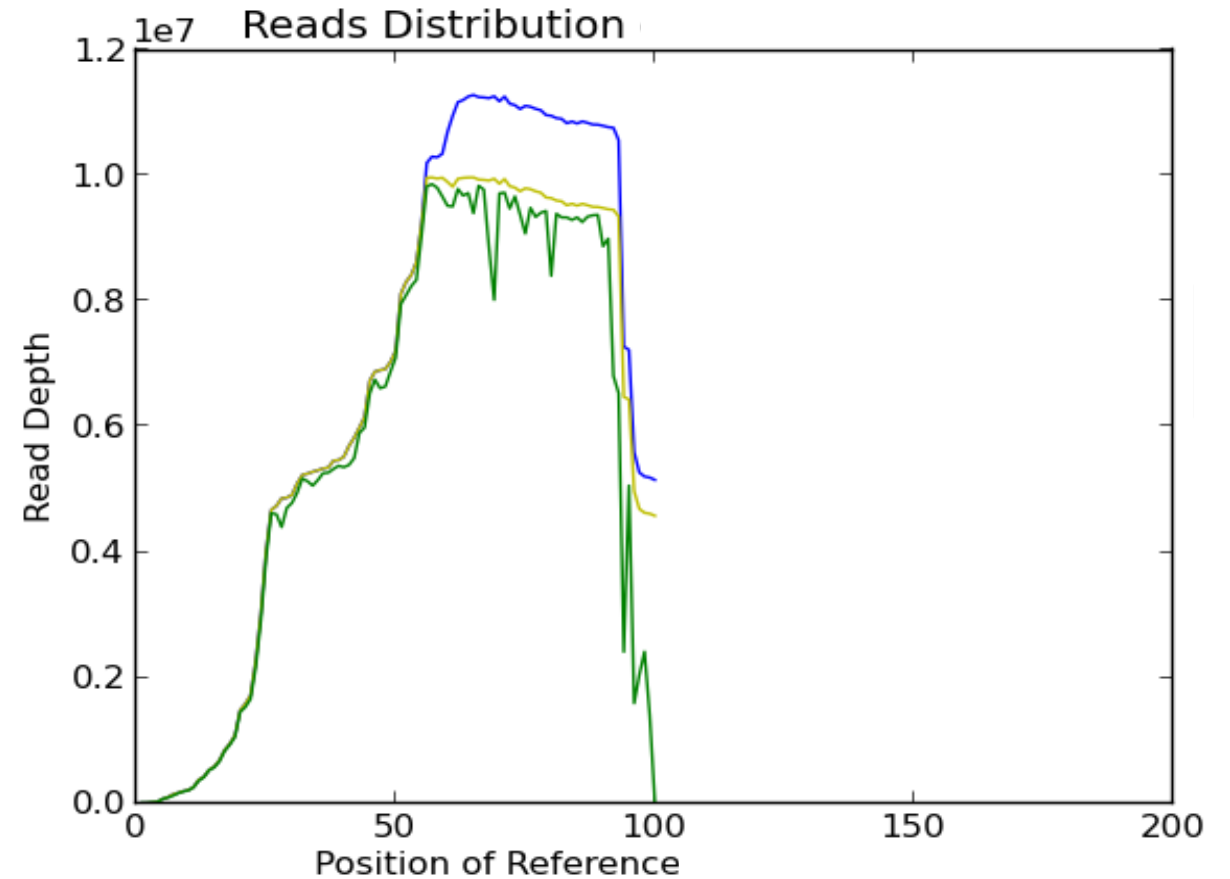
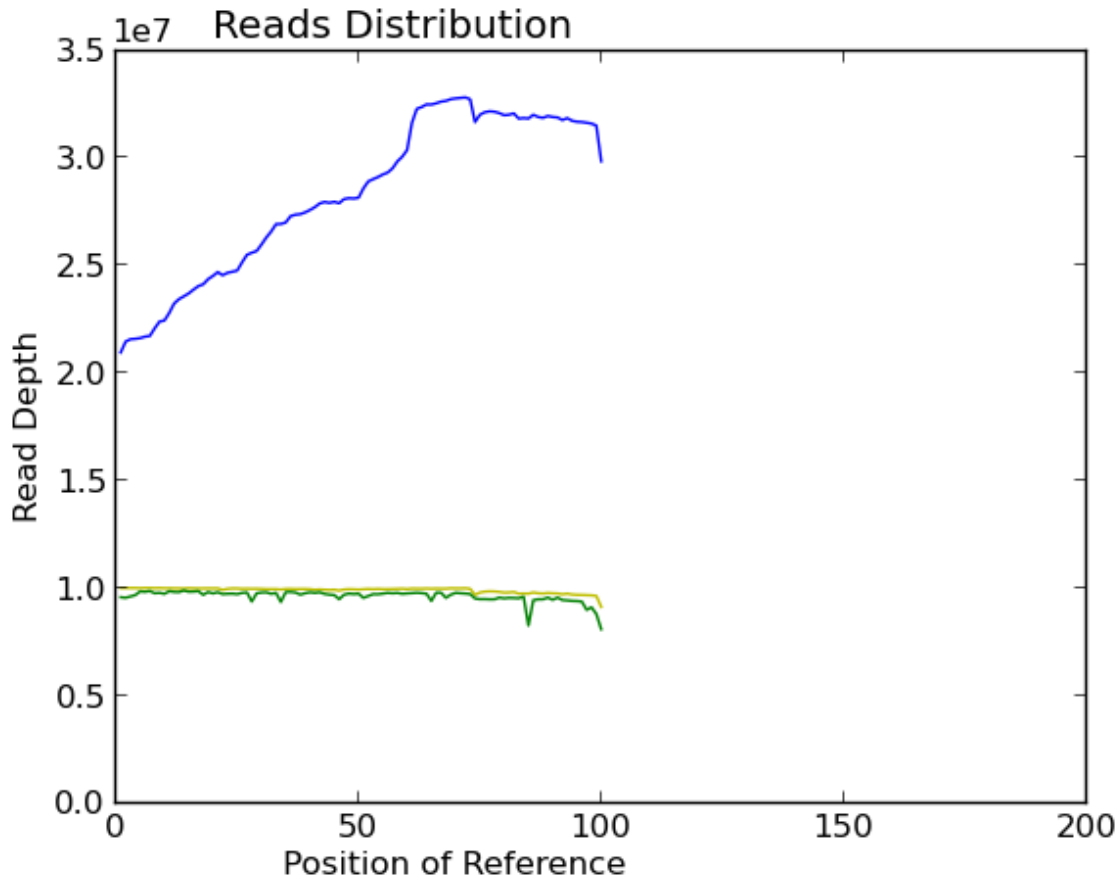


- **Kit B**
- Dilution 1:5
- Average Size: 279 bp

- Data analysis was performed with our proprietary Variant Caller Algorithm (VCA) 2.0
 - No changes made to the algorithm itself

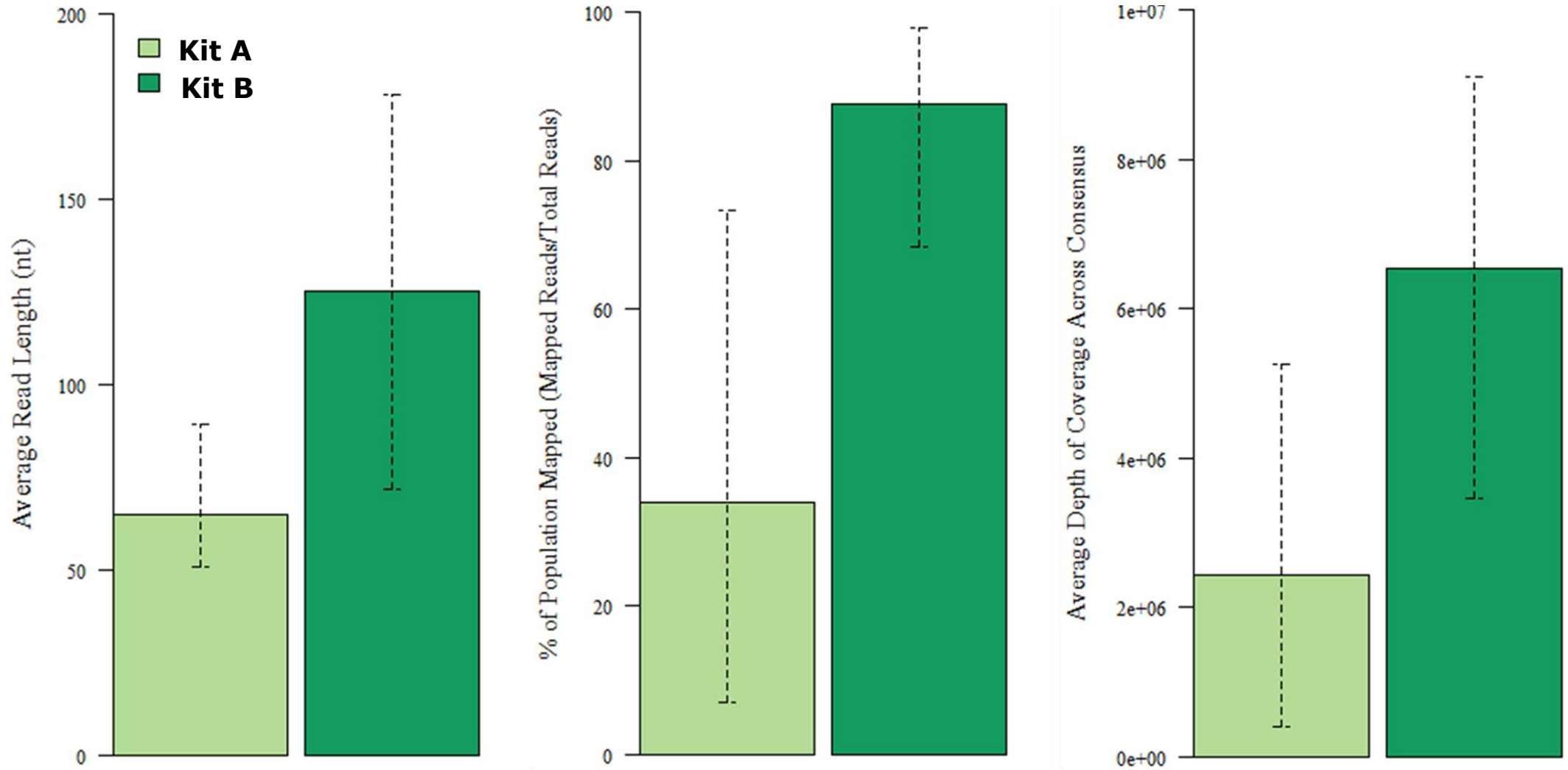
	Kit A	Kit B
Reference Coverage (%)	100	100
Consensus Similarity to Reference (%)	100	100
Total # reads used for mapping	39 million	35 million
% population mapped	30.13	94.5

Read Distribution and Depth – Side by Side



- Alignment Coverage
- Consensus Coverage (DP, raw reads)
- Consensus Coverage (DP4, High Quality Reads)

Kit A vs. Kit B Overview



Key Takeaways

- 1 sgRNA ID by NGS can be performed using Kit B
- 2 NGS provides the level of sensitivity and depth of coverage to accompany current QC technologies
- 3 MilliporeSigma will have a GMP offering starting July 2024!

Learn more:
SigmaAldrich.com/NGS

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